

Making Sense Out of Transcriptome

Integrative Bioinformatic Approaches

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Chapter 1: Interpreting genome-wide expression profiles - A knowledge-based approach

Introduction

Genes typically operate in a sophisticated network of interactions and it is now well recognized that co-expressing genes tend to be playing some common roles in the cell. Recent evidences also suggest functionally related genes map close even in the eukaryotic genomes. Complex phenotypic traits, including diseases are now considered from a systems biology perspective. Thus, there is a clear necessity for methods and tools which can help to understand genome-scale experiments (for e.g. microarray-based gene expression) from a systems biology perspective.

Genome-wide expression analysis with DNA microarrays has become a mainstay of genomics research. In fact, the challenge no longer lies in obtaining gene expression profiles, but rather in interpreting the results to gain insights into biological mechanisms (*Subramanian et al., PNAS, 102: 15545-15550*). A typical experiment generates

mRNA expression profiles for thousands of genes from a collection of samples belonging to different classes. The genes are ordered in a ranked list based on their differential expression between the classes. The proper interpretation of this data requires an integrative systems biology-based functional annotation wherein the collective properties of groups of genes are taken into account rather than individual genes. The principal challenge then is to extract meaning from this list(s) – what is the common or unifying theme(s)?

- Common Regulation (shared cis regulatory elements or transcription factor binding sites)
- Common Biological Function (common pathways or processes)
- Chromosomal Location

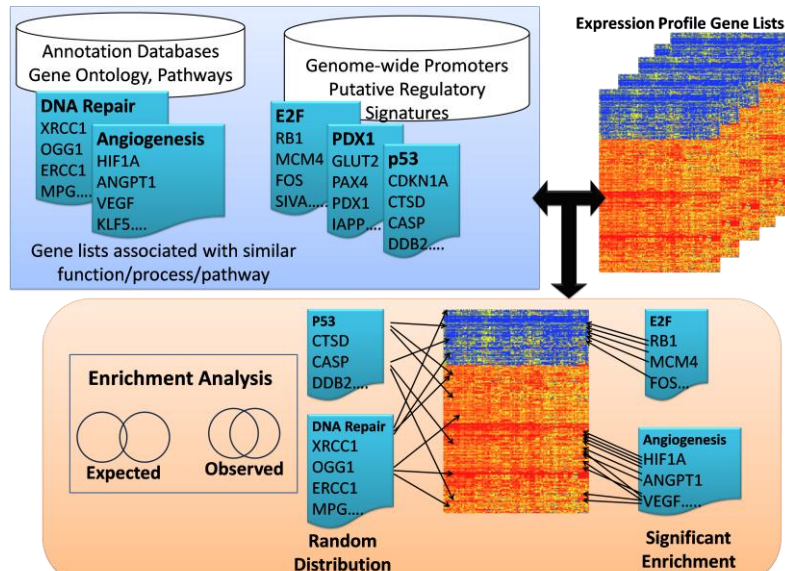


Figure 1: Enrichment analysis for functional and regulatory analysis aimed at identifying specific functions or processes or pathways (GO, KEGG) and transcription factor binding sites that are common for a group of coexpressed genes.

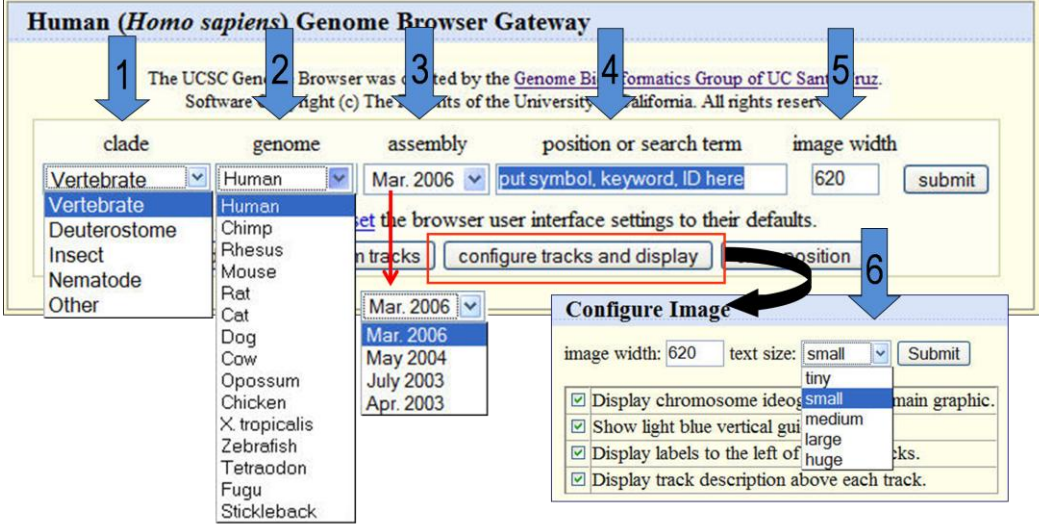
Chapter 2: Strategies for Identifying Putative Gene Regulatory Regions in a Group of Genes

Objectives

- i. Fetch the promoter sequences from a group of coexpressed genes
- ii. Identify the common/shared transcription factor binding sites (TFBSs) or *cis*-elements for this group of promoters
 - a. Known
 1. Non-conserved
 2. Conserved
 - b. Unknown/Novel
 1. Non-conserved
 2. Conserved

Fetching the Promoter Sequence

1. Go to the UCSC genome browser home page (<http://genome.ucsc.edu>) and get the sequence for
 - a. Single gene:
 - i. Using gene symbols:
 - From the home page i.e. <http://genome.ucsc.edu>, click on the “Genomes” link (top navigation bar - left hand corner). Once you are on the Genome Browser Gateway page, select whichever genome you are interested in. In the box under “position or search term” enter the gene symbol and click submit.
 - The following pages list your query results under different categories (e.g. Known Genes, RefSeq Genes, etc.). What this means that your query has results from these different tables or databases.
 - Click on the entries below RefSeq Genes (wherever available; RefSeq or Reference Sequence is a database of curated mRNAs from NCBI) and this will take you to the Genome Browser page.
 - Click on the “DNA” (navigation bar on the top)
 - On the “Get DNA in Window” page, enter your sequence retrieval region options (for e.g. how many base pair upstream, etc.)
 - If you want to mask the repeat regions, check that option under “Sequence Formatting Options”
 - Finally click on “get DNA” to get the sequence in fasta format.
 - Explore the “extended case/color options”
 - ii. Using accession numbers
 - Same as above but using accession number instead of gene symbols.



Human (*Homo sapiens*) Genome Browser Gateway

1 The UCSC Genome Browser was created by the Genome Bioinformatics Group of UC Santa Cruz. Copyright (c) The Regents of the University of California. All rights reserved.

2

3

4

5

6

clade: Vertebrate, Vertebrate, Deuterostome, Insect, Nematode, Other

genome: Human, Chimp, Rhesus, Mouse, Rat, Cat, Dog, Cow, Opossum, Chicken, X. tropicalis, Zebrafish, Tetraodon, Fugu, Stickleback

assembly: Mar. 2006, Mar. 2006, May 2004, July 2003, Apr. 2003

position or search term: put symbol, keyword, ID here

image width: 620

submit

set the browser user interface settings to their defaults.

configure tracks and display

position

Configure Image

image width: 620 text size: small, tiny, medium, large, huge

Submit

☒ Display chromosome ideograms in main graphic.

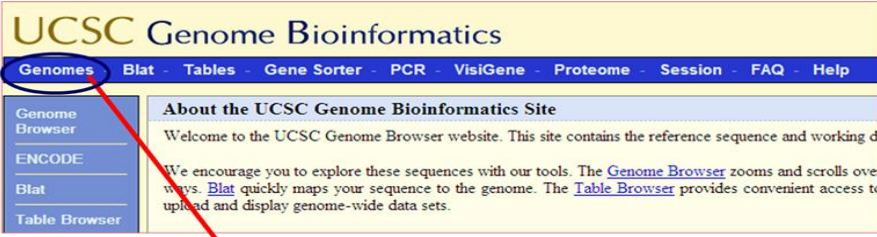
☒ Show light blue vertical guide tracks.

☒ Display labels to the left of tracks.

☒ Display track description above each track.

Genome Browser Gateway choices:

1. Select Clade
2. Select genome/species: You can search only one species at a time
3. Assembly: the official backbone DNA sequence
4. Position: location in the genome to examine or search term (gene symbol, accession number, etc.)
5. Image width: how many pixels in display window; 5000 max
6. Configure: make fonts bigger + other options



UCSC Genome Bioinformatics

Genomes | Blat | Tables | Gene Sorter | PCR | VisiGene | Proteome | Session | FAQ | Help

Genome Browser

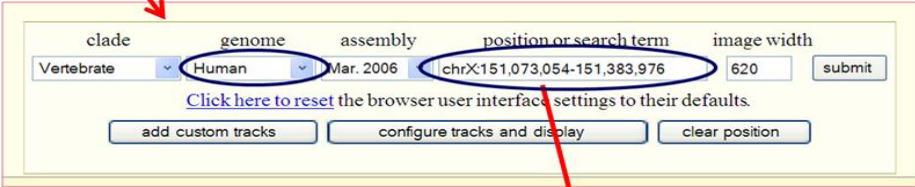
ENCODE

Blat

Table Browser

About the UCSC Genome Bioinformatics Site

Welcome to the UCSC Genome Browser website. This site contains the reference sequence and working data sets for the human genome. We encourage you to explore these sequences with our tools. The [Genome Browser](#) zooms and scrolls over the genome. [Blat](#) quickly maps your sequence to the genome. The [Table Browser](#) provides convenient access to upload and display genome-wide data sets.



clade: Vertebrate

genome: Human

assembly: Mar. 2006

position or search term: chrX:151,073,054-151,383,976

image width: 620

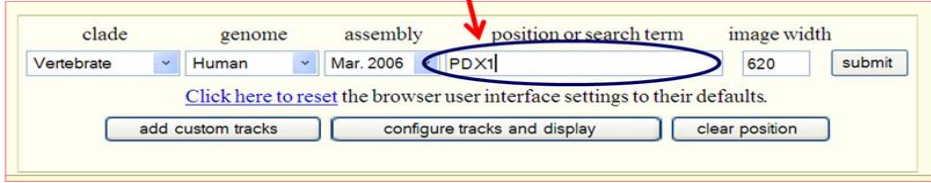
submit

[Click here to reset](#) the browser user interface settings to their defaults.

add custom tracks

configure tracks and display

clear position



clade: Vertebrate

genome: Human

assembly: Mar. 2006

position or search term: PDX1

image width: 620

submit

[Click here to reset](#) the browser user interface settings to their defaults.

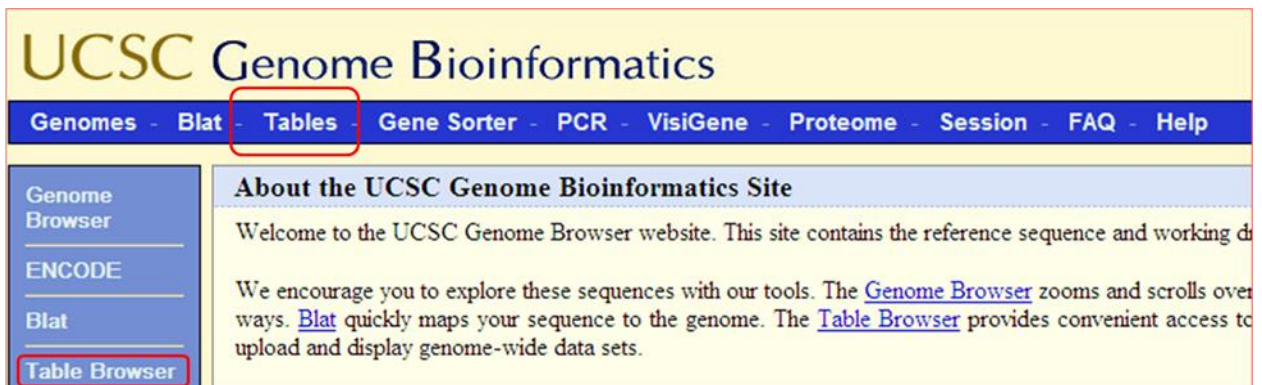
add custom tracks

configure tracks and display

clear position

- Use the option “RefGene” instead of “RefFlat”

2. **Advanced:** What if you want to download upstream 1 kb **plus** 200 bp downstream of the transcription start site? It can be done but you need to get the start positions of all your genes of interest and then (using excel) start subtracting 1000 from the start and also start adding 200. Thus, you will get a new start and end and you can use these to fetch the genomic sequences of desired length.
3. **Advanced:** How can I get a list of all SNPs occurring in the upstream 1 kb regions of genes of my interest?
 - a. To do this you first need the coordinates or positions of the upstream regions (1 kb regions' start and end positions) (see 2 above)
 - b. Then using these coordinates or regions as the base you can intersect with other features (for e.g. get me all SNPs that intersect or fall within these selected regions. Use the "intersection" feature from the table browser options.
4. **LIMITATIONS:**
 - a. Transcription Start Sites (TSS) are still not well defined. Surprisingly the experimentally validated promoters for human and mouse are only about 1870 and 200 respectively! The **Eukaryotic Promoter Database (EPD)** is an annotated non-redundant collection of eukaryotic POL II promoters, for which the transcription start site has been determined experimentally. EPD can be accessed at <http://www.epd.isb-sib.ch/>. For all practical purposes, we consider the region upstream to the NCBI's RefSeq mRNAs (<http://www.ncbi.nlm.nih.gov/RefSeq/>) as putative promoters. There is no single, perfect TSS or promoter prediction algorithms. Of the available ones, the firstEF (first exon finder; <http://rulai.cshl.edu/tools/FirstEF/>) does a reasonably good job (tested using experimentally validated promoters).
 - b. Regulatory regions (especially enhancers and silencers) can occur just anywhere (intronic, downstream, farther upstream, UTRs).
 - c. Regulatory mechanisms other than transcriptional: Posttranscriptional (e.g. miRNAs), posttranslational (protein modifications) or epigenetic mechanisms (differential effects of chromosome or chromatin packaging rather than by differences in DNA sequence).



UCSC Genome Bioinformatics

Genomes - Blat - **Tables** - Gene Sorter - PCR - VisiGene - Proteome - Session - FAQ - Help

Genome Browser
 ENCODE
 Blat
Table Browser

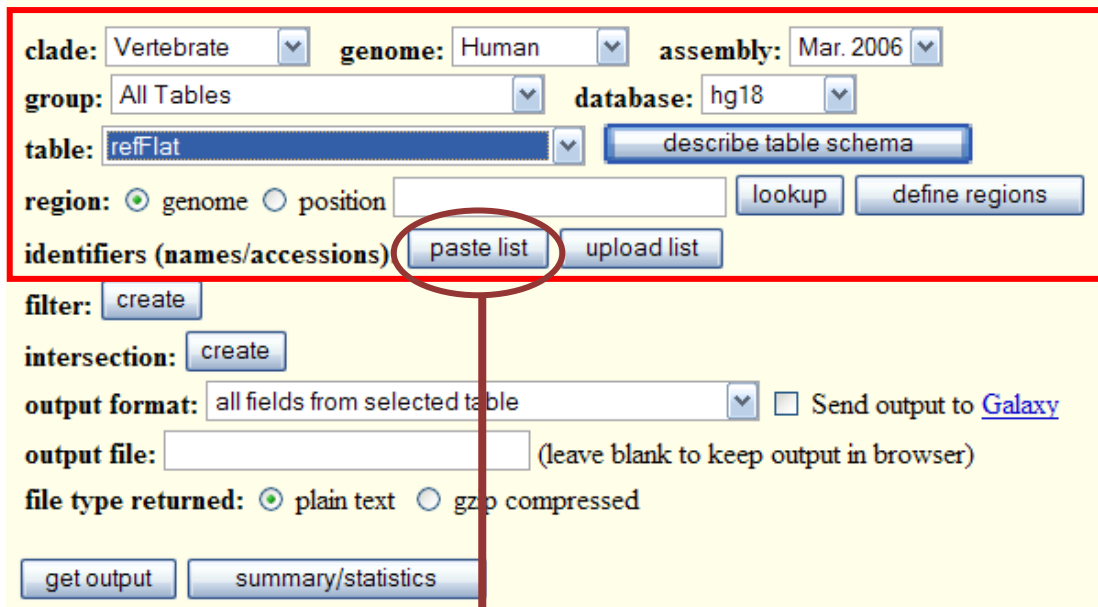
About the UCSC Genome Bioinformatics Site

Welcome to the UCSC Genome Browser website. This site contains the reference sequence and working d

We encourage you to explore these sequences with our tools. The [Genome Browser](#) zooms and scrolls over ways. [Blat](#) quickly maps your sequence to the genome. The [Table Browser](#) provides convenient access to upload and display genome-wide data sets.

Table Browser

Use this program to retrieve the data associated with a track in text format, to calculate intersection: application see [Using the Table Browser](#) for a description of the controls in this form, the [User's Guide](#) narrated presentation of the software features and usage. For more complex queries, you may want usage restrictions associated with these data.



clade: Vertebrate genome: Human assembly: Mar. 2006

group: All Tables database: hg18

table: refFlat describe table schema

region: ☒ genome ☐ position lookup define regions

identifiers (names/accessions) paste list upload list

filter: create

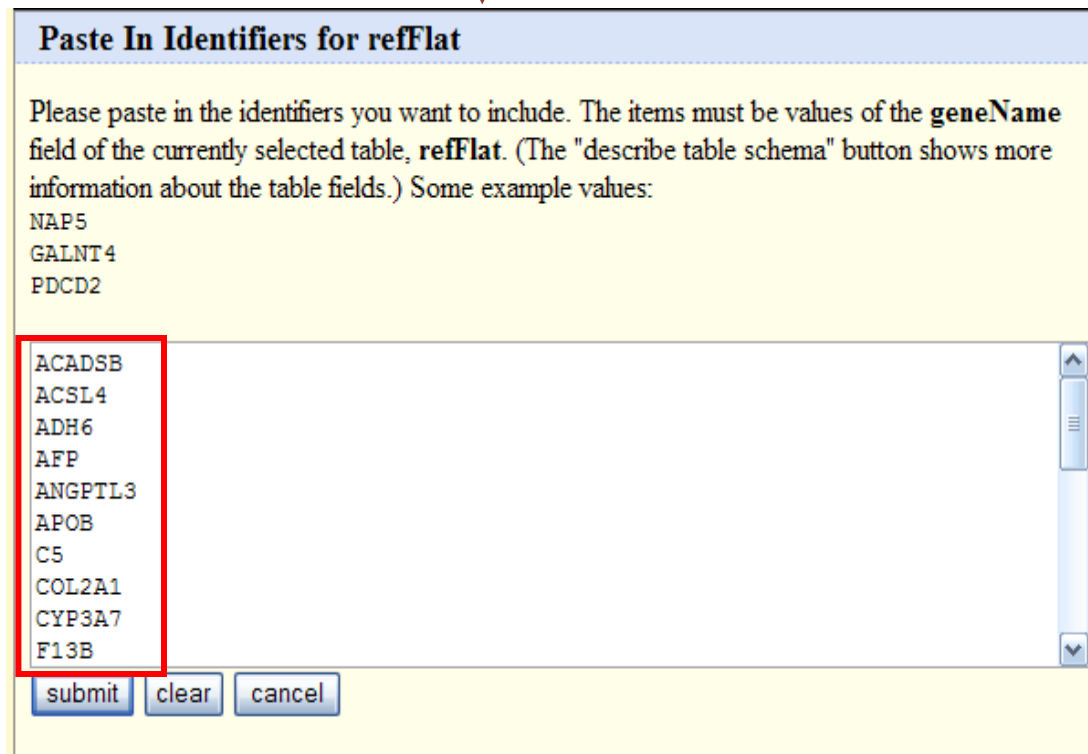
intersection: create

output format: all fields from selected table ☐ Send output to [Galaxy](#)

output file: (leave blank to keep output in browser)

file type returned: ☒ plain text ☐ gzip compressed

get output summary/statistics



Paste In Identifiers for refFlat

Please paste in the identifiers you want to include. The items must be values of the **geneName** field of the currently selected table, **refFlat**. (The "describe table schema" button shows more information about the table fields.) Some example values:

NAP5
GALNT4
PDCD2

ACADSB
ACSL4
ADH6
AFP
ANGPTL3
APOB
C5
COL2A1
CYP3A7
F13B

submit clear cancel

Table Browser (Input Identifiers)

Use this program to retrieve the data associated with a track in text format, to calculate intersection application see [Using the Table Browser](#) for a description of the controls in this form, the [User's C](#) narrated presentation of the software features and usage. For more complex queries, you may want to use restrictions associated with these data.

clade: genome: assembly:

group: database:

table:

region: ☒ genome ☐ position

identifiers (names/accessions):

filter:

intersection:

output format: ☐ Send output to [Galaxy](#)

output file: (leave blank to keep output in browser)

file type returned: ☒ plain text ☐ gzip compressed

[illegible]

Home Genomes Genome Browser Blat Tables Gene Sorter PCR Session FAQ Help

Table Browser

Use this program to retrieve the data associated with a track in text format, to calculate intersections between tracks, and to retrieve DNA sequence covered by a track. For help in using this application see [Using the Table Browser](#) for a description of the controls in this form, the [User's Guide](#) for general information and sample queries, and the [OpenHelix Table Browser tutorial](#) for a narrated presentation of the software features and usage. For more complex queries, you may want to use [Galaxy](#) or our [public MySQL server](#). Refer to the [Credits](#) page for the list of contributors and usage restrictions associated with these data.

clade: Vertebrate genome: Human assembly: Mar 2006

group: All Tables database: hg18

table: refFlat describe table schema

region: ☒ genome ☐ position chrX:151073054-151383976 lookup define regions

identifiers (names/accessions): paste list upload list

filter: create

intersection with UserTrack: edit clear

output format: all fields from selected table ☐ Send

output file: (leave blank to keep output in browser)

file type returned: ☒ plain text ☐ gzip compressed

Note: Intersection doesn't work with all fields or selected fields output.
get output summary/statistics

To reset all user cart settings (including custom tracks), [click here](#).

1. Select "refFlat" under "table"
2. Ensure that "region" is "genome"
3. Click on "paste list"

Home Genomes Genome Browser Blat Tables Gene Sorter PCR Session FAQ Help

Paste In Identifiers for refFlat

Please paste in the identifiers you want to include. The items must be values of the `geneName` field of the currently selected table, `refFlat`. (The "describe table schema" button shows more information about the table fields.) Some example values:

1. Paste the gene symbols

2. Remember it is case-sensitive:

- Human: all upper case (e.g. XRCC1)
- Mouse: lower case (first letter upper case. E.g. Xrcc1)

ABCC3
ABHD2
ACT1
ADH1C
AFOS
AFOS4
AGP9
ASL
AZGP1

submit clear cancel

Home Genomes Genome Browser Blat Tables Gene Sorter PCR Session FAQ Help

Output refFlat as Custom Track

Custom track header:

name: UpSL1kb

description: table browser query on refFlat

visibility: pack

url: (leave blank to keep output in browser)

Create one BED record per:

☒ Whole Gene

☒ Upstream by 1000 bases

☐ Exons plus 0 bases at each end

☐ Introns plus 0 bases at each end

☐ 5' UTR Exons

☐ Coding Exons

☐ 3' UTR Exons

☐ Downstream by 200 bases

Note: If a feature is close to the beginning or end of a chromosome and upstream/downstream bases are added, they will be truncated in order to avoid extending past the edge of the chromosome.

get custom track in table browser get custom track in file

get custom track in genome browser cancel

Enter number of bp you want to analyze/download

Home Genomes Genome Browser Blat Tables Gene Sorter PCR Session FAQ Help

Table Browser

Use this program to retrieve the data associated with a track in text format, to calculate intersections between tracks, and to retrieve DNA sequence covered by a track. For help in using this application see [Using the Table Browser](#) for a description of the controls in this form, the [User's Guide](#) for general information and sample queries, and the [OpenHelix Table Browser tutorial](#) for a narrated presentation of the software features and usage. For more complex queries, you may want to use [Galaxy](#) or our [public MySQL server](#). Refer to the [Credits](#) page for the list of contributors and usage restrictions associated with these data.

clade: Vertebrate genome: Human assembly: Mar 2006

group: All Tables database: hg18

table: refFlat describe table schema

region: ☒ genome ☐ position chrX:151073054-151383976 lookup define regions

identifiers (names/accessions): paste list upload list clear list

filter: create

intersection: create

output format: custom track ☐ Send output to Galaxy

output file: (leave blank to keep output in browser)

file type returned: ☒ plain text ☐ gzip compressed

get output summary/statistics

To reset all user cart settings (including custom tracks), [click here](#).

Select the output format as "custom track"

One drawback with this output is it doesn't tell you which SNPs are in the upstream region of which gene. However, since the positions of SNPs are included, you can compare them with the gene coordinates and figure it out.

Identification of Putative Common/Shared *Cis*-Elements

Known

Non-conserved

GEMS Launcher (Genomatix) Search for common TF sites in multiple sequences

URL: <http://www.genomatix.de>

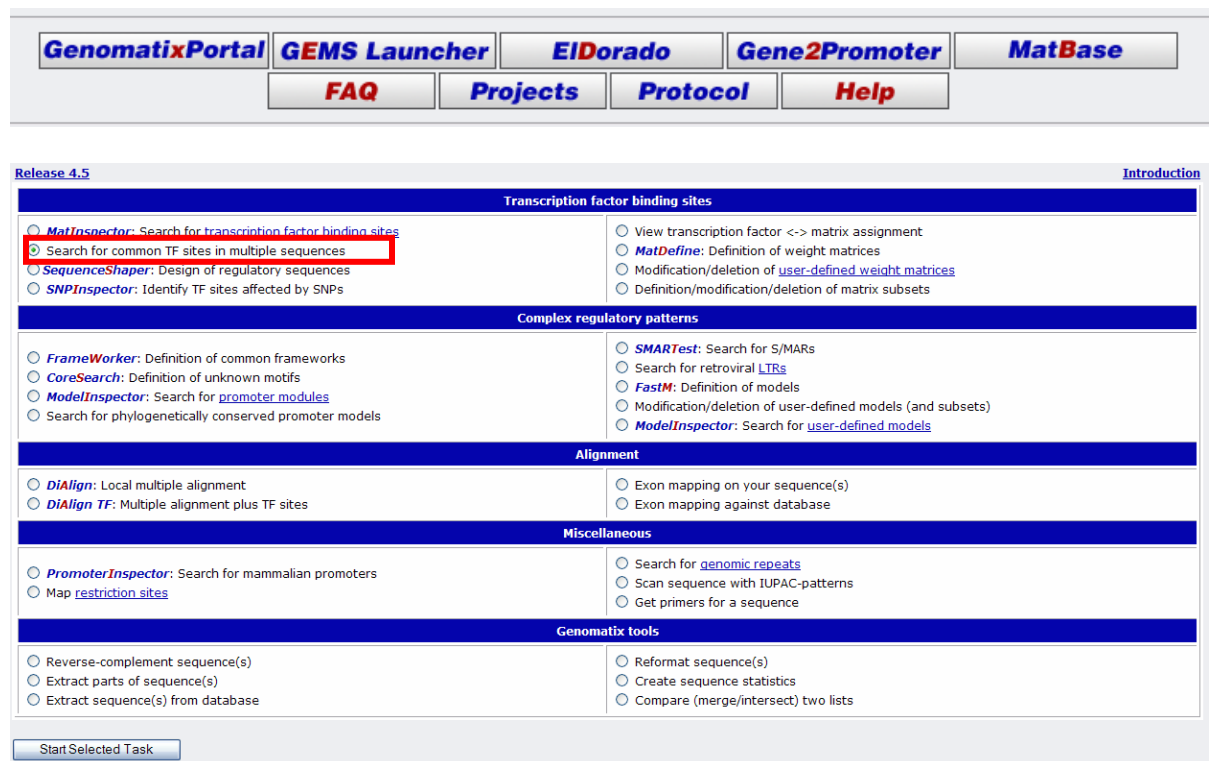
Access: Licensed software (free for 20 analyses per month).

Description/Utility: You can search for transcription factor binding sites (TFBS) that are common to all or a subset of your input sequences. The results are displayed graphically. You can enter the percentage of sequences that have to contain the common sites. The search is based on known TFBSs that are stored as position weight matrices (PWMs) in the library.

Input: Fasta sequences (max 100 sequences) or any other format.

Output: Graphical and tab-delimited/spreadsheets

Other Comments: Genomatix has several other useful applications/tools that help to understand the molecular mechanisms of gene regulation as a central part of systems biology. The MatInspector tool of Genomatix is one of the widely used tools to identify potential binding sites in a DNA sequence.



The screenshot shows the Genomatix Portal interface. At the top, there are navigation buttons: GenomatixPortal, GEMS Launcher, EIDorado, Gene2Promoter, MatBase, FAQ, Projects, Protocol, and Help. Below this, the main content area is titled 'Transcription factor binding sites' and lists several tools with radio button selection options. The tool 'Search for common TF sites in multiple sequences' is highlighted with a red box. Other tools include MatInspector, SequenceShaper, SNPIInspector, FrameWorker, CoreSearch, ModelInspector, SMARTest, Search for retroviral LTRs, FastM, and ModelInspector. The interface also includes sections for 'Complex regulatory patterns', 'Alignment', 'Miscellaneous', and 'Genomatix tools'.

Release 4.5 Introduction

Transcription factor binding sites

- ☐ MatInspector: Search for [transcription factor binding sites](#)
- ☒ Search for common TF sites in multiple sequences
- ☐ SequenceShaper: Design of regulatory sequences
- ☐ SNPIInspector: Identify TF sites affected by SNPs
- ☐ View transcription factor <-> matrix assignment
- ☐ MatDefine: Definition of weight matrices
- ☐ Modification/deletion of [user-defined weight matrices](#)
- ☐ Definition/modification/deletion of matrix subsets

Complex regulatory patterns

- ☐ FrameWorker: Definition of common frameworks
- ☐ CoreSearch: Definition of unknown motifs
- ☐ ModelInspector: Search for [promoter modules](#)
- ☐ Search for phylogenetically conserved promoter models
- ☐ SMARTest: Search for S/MARs
- ☐ Search for retroviral [LTRs](#)
- ☐ FastM: Definition of models
- ☐ Modification/deletion of user-defined models (and subsets)
- ☐ ModelInspector: Search for [user-defined models](#)

Alignment

- ☐ DiAlign: Local multiple alignment
- ☐ DiAlign TF: Multiple alignment plus TF sites
- ☐ Exon mapping on your sequence(s)
- ☐ Exon mapping against database

Miscellaneous

- ☐ PromoterInspector: Search for mammalian promoters
- ☐ Map [restriction sites](#)
- ☐ Search for [genomic repeats](#)
- ☐ Scan sequence with IUPAC-patterns
- ☐ Get primers for a sequence

Genomatix tools

- ☐ Reverse-complement sequence(s)
- ☐ Extract parts of sequence(s)
- ☐ Extract sequence(s) from database
- ☐ Reformat sequence(s)
- ☐ Create sequence statistics
- ☐ Compare (merge/intersect) two lists

Start Selected Task

☐ Choose from your previously uploaded sequences

☒ or enter the correctly formatted DNA sequence(s)

☐ or upload a file containing sequence(s) (max. 100 MB)

☐ or enter accession number(s)

Sorry, no uploaded sequences yet!

Supported formats: [plain](#), [EMBL](#), [FASTA](#), [GCG/RSF](#), [GenBank](#), [IG](#)

```
>Upst_ANGPTL3 range=chr1:62834775-62835774
5'pad=0 3'pad=0 revComp=FALSE strand=+
repeatMasking=lower
aattgctgggctcacgcttgtaaatcggtgggctcatgctgtga
aattt
```

Name for your sequence file:

Optional name for your sequence-file on the server:

(separated by spaces or commas)

Library Selection

Please select one of the following libraries:

Transcription factor binding sites (Weight matrices)

Plant IUPAC library (based on PLACE)

- See the [list](#) of available weight matrices
- See the [list](#) of available IUPAC library strings

Matrix Parameters		Less options
Library version	Matrix Library 6.3	
Matrix group (View transcription factor <-> matrix assignment)	<input type="checkbox"/> Fungi <input type="checkbox"/> Other Functional Elements <input type="checkbox"/> Insects <input type="checkbox"/> Plants <input type="checkbox"/> Miscellaneous <input checked="" type="checkbox"/> Vertebrates	
Matrix families	<ul style="list-style-type: none"> - use all matrices from selected groups - use previously defined matrix subsets - continue with subset definition from selected groups 	
Matrix filters (only available for vertebrates)	<ul style="list-style-type: none"> - matches to matrix families (see matrix families) - matches to individual matrices (see matrices) <p>Sorry, tissue filtering is ONLY available for licensed users of MatInspector! (More info...)</p> <p>If you want to use tissue filtering please order unlimited access to MatInspector.</p>	
Core similarity	0.85	
Matrix similarity	Optimized	
TF sites common to	2 of 33 (6 %) of input sequences	
Result name		
Result name (optional)	<input type="text"/> (special characters like "#\$%&+./:;<=>?@ not allowed)	
<input type="button" value="Submit Query"/> <input type="button" value="Reset Form"/>		



Family/Matrix	p-value	#sequences	UpSt_ANGPTL3	UpSt_F13B	UpSt_APOB	UpSt_TM4SF4	UpSt_SLC2A2
V\$HOMF/V\$NOBOX.01	1.08E-45	25	1	1	0	3	3
V\$HOMF/V\$DLX1.01	2.44E-30	25	3	2	0	4	3
V\$HOMF/V\$DLX3.01	1.62E-19	24	1	2	0	4	2
V\$GATA/V\$GATA1.05	1.51E-16	21	1	3	0	0	4
V\$HOXF/V\$PCE1.01	3.40E-16	23	1	1	0	3	1
V\$GATA/V\$GATA1.04	1.18E-14	22	1	2	0	1	3
V\$GATA/V\$GATA2.01	3.82E-14	22	1	3	0	0	4
V\$GATA/V\$GATA.01	8.30E-14	25	0	0	0	0	2
V\$MYBL/V\$VMYB.03	1.58E-13	10	0	0	0	0	0
V\$GATA/V\$GATA2.02	5.72E-13	21	1	1	0	1	3
V\$GATA/V\$GATA3.01	5.80E-13	18	1	2	0	0	4
V\$SORY/V\$SRY.01	2.66E-12	17	3	1	0	1	2
V\$FKHD/V\$HFH1.01	1.73E-11	20	1	1	0	1	1
V\$HOXF/V\$PHOX2.01	3.07E-11	24	0	3	1	2	2
V\$TBPF/V\$LTATA.01	7.50E-11	25	4	3	0	0	4
V\$EV11/V\$MEL1.01	9.99E-11	15	0	1	0	2	1

Conserved

CisMols

URL: <http://cismols.cchmc.org>

Access: Free; Web-based

Description/Utility: Filtering candidate transcription factor binding site clusters (cis-regulatory element clusters) based on sequence conservation is helpful for an individual ortholog gene pair, but combining data from cis-conservation and coordinate expression across multiple genes is a more difficult problem. To approach this, we have extended an ortholog gene pair database with additional analytical architecture to allow for the analysis and identification of maximal numbers of compositionally similar and phylogenetically conserved cis-regulatory element clusters from a list of user-selected genes. Starting with identification of cis-clusters in phylogenetic footprints, we intend to extend the query to identify compositionally similar cis regulatory element clusters that occur in groups of co-regulated genes within each of their ortholog-pair evolutionarily conserved cis-regulatory regions. These computationally predicted cis-clusters, which we call as cismols, could serve as valuable probes for genome wide identification of regulatory regions and novel gene targets.


Support: Please refer to the “Help” section

(<http://info.cchmc.org/help/cismols/index.html>) on CisMols home page

(<http://cismols.cchmc.org>). For accounts, any problems or questions or analysis, send a mail to anil.jegga@cchmc.org.

Input: Human or mouse gene symbols or RefSeq accession numbers (hint: start with NM_).

Output: Graphical output (can be stored as pdf or any other image formats). Data can be downloaded in a spreadsheet.



CisMols

View Computationally identified *Cis*-regulatory modules, called CisMols, that occur in groups of Coexpressed or Related genes within their ortholog-pair evolutionarily conserved *cis*-regulatory regions.

[Logout](#)
[Home](#)
[Help](#)

You need to have a login account; contact anil.jegga@cchmc.org

Choose Data Source:
☒ Pipeline Genes ☐ Manually Curated Genes

CisMols Analyzer **1**

Welcome back, ajegga!

- [Create Gene List](#)
Create Gene List.
- [Search for Genelist](#)
Search for specific Genelists.
- [Compare Two Genelists](#)
Statistical Comparison of clusters in two genelists.
- [Projects](#)
Add, remove, or modify projects.

Administrative Features

- [Users](#)
Add, remove, or modify CisMols users.

2

Search for Genes to Add to List

Comma or Space Delimited, Case Insensitive Search.

Accession Number: Contained in.

Gene Symbol: ACADSB, ACSL4, ADHE HGNC/MGI Approved; Exact match

Description: Contained in.

Sequence Group: All Groups

3

Select Genes for Finding CisMols

Enter length in base pairs and choose region to modify the default values of 10kb up and down stream of the gene.

Step 1. Specify No. of Base Pairs: 1000

Step 2. Select Region with respect to: ☒ First Exon ☐ Last Exon ☐ Gene

Step 3. Click on symbol to populate: ☒ Down Stream ☐ Up Stream ☐ Up and Down Stream

☒ Select All Genes

Select	Accession Number	Base Sequence Name Ortholog Sequence Name	First Exon	Last Exon	Diff. btwn Base and Ortholog exon counts	From	To
<input checked="" type="checkbox"/>	hgNM_001609	ACADSB human Acyl-Coenzyme A dehydrogenase, short/	40001	89272	-1	39001	40001
	mgNM_025826	Acadsb mouse Acyl-Coenzyme A dehydrogenase, short/	40001	76656			
<input checked="" type="checkbox"/>	hgNM_022977	ACSL4 human Acyl-CoA synthetase long-chain family	40001	132054	1	39001	40001
	mgNM_019477	Acs4 mouse Acyl-CoA synthetase long-chain family	40001	110451			
<input checked="" type="checkbox"/>	hgNM_001134	AFP human Alpha-fetoprotein chr4	40001	59560	0	39001	40001
	mgNM_007423	Afp mouse Alpha fetoprotein chr5	40001	58170			
<input checked="" type="checkbox"/>	hgNM_014495	ANGPTL3 human Angiopoietin-like 3 chr1	40001	47994	0	39001	40001
	mgNM_013913	Angptl3 mouse Angiopoietin-like 3 chr4	40001	47037			

4

Search for Genes to Add to List 5

Comma or Space Delimited, Case Insensitive Search.

Accession Number:

Gene Symbol:

Description:

Sequence Group:

Contained in.

HGN/NCBI Approved, Exact match

Contained in.

Gene Cart Update or Submit 6

Associate to New Project:

Associate to Existing Project:

Gene List Name:

Description:

Email Address:

Visibility:

Create Project

DHC_Workshop

Fetal_Liver_18

UpSt 1 kb

anil.jegga@cchmc.org

☐ Private ☒ Public

Submit Gene List

Number of genes in cart: 18

	ORTHOLOG GENE PAIR NAMES (Select to remove)	EXON START	EXON END	FROM	TO
<input type="checkbox"/>	ACSL4 human Acyl-CoA synthetase long-chain family	40001	132054	39001	40001
	Acs14 mouse Acyl-CoA synthetase long-chain family	40001	110451		
<input type="checkbox"/>	SERPINA7 human Serpin peptidase inhibitor, clade A	40001	43870	39001	40001
	Serpina7 mouse Serine (or cysteine) peptidase inhi	40001	43544		
<input type="checkbox"/>	SLC2A2 human Solute carrier family 2 (facilitated	40001	70632	39001	40001
	Slc2a2 mouse Solute carrier family 2 (facilitated	40001	70351		
<input type="checkbox"/>	PANK1 human Pantothenate kinase 1 chr10	40001	102467	39001	40001
	Pank1 mouse Pantothenate kinase 1 chr19	40001	107023		

Welcome back, ajegga! 7

- [Create Gene List](#)
Create Gene List.
- [Search for Genelist](#)
Search for specific Genelists.
- [Compare Two Genelists](#)
Statistical Comparison of clusters in two genelists.
- [Projects](#)
Add, remove, or modify projects.

Administrative Features

- [Users](#)
Add, remove, or modify CisMols users.

Gene List Search 8

View Gene Lists 9

Search Criteria

Gene Accession Number: Contained in

Gene Symbol: HGNC/MGI Approved, Exact match

Gene List Name: Contained in.

Project:

Researcher:


Gene List has at least: Clusters

Gene List has at least: Genes

Search with default values to view all genelists.

[Show Public](#) ☒ YES ☐ NO

My Gene Lists

Gene List Name	Project Name	List Description	No. of Genes	No. of Clusters	Created By	Created On	Delete
Fetal Liver 18	DHC Workshop	UpSt 1 kb	18	2731	ajegga	2007-09-18	

Edit Gene List Properties 10

Name: ☒ Public ☐ Private

Genes

Accession Number	Base Sequence Name Ortholog Sequence Name	First Exon	Last Exon	Clustering Start	Clustering End	Difference in Base and Ortholog Exons
hgNM_001994	F13B human Coagulation factor XIII, B polypeptide	40001	68044	39001	40001	0
mgNM_031164	F13b mouse Coagulation factor XIII, beta subunit c	40001	62030	39471	40003	
hgNM_000508	FGA human Fibrinogen alpha chain chr4	40001	47586	39001	40001	1
mgNM_010196	Fga mouse Fibrinogen, alpha polypeptide chr3	40001	46103	39050	39968	
hgNM_014495	ANGPTL3 human Angiopoietin-like 3 chr1	40001	47994	39001	40001	0
mgNM_013913	Angptl3 mouse Angiopoietin-like 3 chr4	40001	47037	39119	39943	
hgNM_001609	ACADSB human Acyl-Coenzyme A dehydrogenase, short/	40001	89272	39001	40001	-1
mgNM_025826	Acad5b mouse Acyl-Coenzyme A dehydrogenase, short/	40001	76656	39084	39967	
hgNM_002108	HAL human Histidine ammonia-lyase chr12	40001	62930	39001	40001	1
mgNM_010401	Hal mouse Histidine ammonia lyase chr10	40001	67976	39197	39993	
hgNM_001844	COL2A1 human Collagen, type II, alpha 1 (primary o	40001	71511	39001	40001	2
mgNM_031163	Col2a1 mouse Procollagen, type II, alpha 1 chr15	40001	68452	38930	39823	
hgNM_005141	FGB human Fibrinogen beta chain chr4	40001	48074	39001	40001	0
mgNM_181849	Fgb mouse Fibrinogen, B beta polypeptide chr3	40001	47484	38903	39973	
hgNM_148977	PANK1 human Pantothenate kinase 1 chr10	40001	102467	39001	40001	0
mgNM_023792	Pank1 mouse Pantothenate kinase 1 chr19	40001	107023	37497	38572	
hgNM_000612	IGF2 human Insulin-like growth factor 2 (somatomed	40001	46047	39001	40001	0
mgNM_010514	Igf2 mouse Insulin-like growth factor 2 chr7	40001	51092	41989	42947	
hgNM_000253	MTTP human Microsomal triglyceride transfer protei	40001	88646	39001	40001	0
mgNM_008642	Mttp mouse Microsomal triglyceride transfer protei	40001	80439	38606	39933	
hgNM_022977	ACSL4 human Acyl-CoA synthetase long-chain family	40001	132054	39001	40001	1
mgNM_019477	Acs14 mouse Acyl-CoA synthetase long-chain family	40001	110451	39032	39871	
hgNM_000340	SLC2A2 human Solute carrier family 2 (facilitated	40001	70632	39001	40001	0
mgNM_031197	Slc2a2 mouse Solute carrier family 2 (facilitated	40001	70351	39328	39791	
hgNM_001134	AFP human Alpha-fetoprotein chr4	40001	59560	39001	40001	0
mgNM_007423	Afp mouse Alpha-fetoprotein chr5	40001	58170	38846	39976	
hgNM_000531	OTC human Ornithine carbamoyltransferase chrX	40001	109252	39001	40001	0
mgNM_008769	Otc mouse Ornithine transcarbamylase chrX	40001	108666	38825	39454	
hgNM_001735	C5 human Complement component 5 chr9	40001	137940	39001	40001	0
mgNM_010406	Hc mouse Hemolytic complement chr2	40001	118066	39257	39949	
hgNM_004617	TM4SF4 human Transmembrane 4 L six family member 4	40001	68635	39001	40001	0
mgNM_145539	Tm4sf4 mouse Transmembrane 4 superfamily member 4	40001	56207	39022	39910	
hgNM_002216	ITIH2 human Inter-alpha (globulin) inhibitor H2 ch	40001	86146	39001	40001	0
mgNM_010582	Itih2 mouse Inter-alpha trypsin inhibitor, heavy c	40001	76073	39255	40031	
hgNM_000354	SERPINA7 human Serpin peptidase inhibitor, clade A	40001	43870	39001	40001	0
mgNM_177920	Serpina7 mouse Serpin peptidase inhibitor, clade A	40001	43544	39212	39974	

CisMols Search

11

Fetal_Liver_18

Min Genes and Sites in a Clusters

Min Genes in Cluster:

Min Sites in Cluster:

Order (top 100 by default) clusters by

☒ No. of Genes

☐ No. of Sites

View Region (base pairs)

From:

To:

[CisMols Search](#)
[Saved Searches](#)
[Clear Form](#)

Select individual sites

And	Or	Not	Site	And	Or	Not	Site
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSAARF	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSAHRR
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSAIRE	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSAP1F
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSAP1R	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSAP2F
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSAP4R	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSAREB
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSARID	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSARP1
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSATBF	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSBARB
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSBC16	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSBEL1
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSBNCF	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSBRAC
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSBRNF	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSBTBF
<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSCABL	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	VSCART

☐ AND

☒ OR

☐ **Select from known Modules**

<input type="checkbox"/> VSAHRR VSSP1F	<input type="checkbox"/> VSETSF VSETSF
<input type="checkbox"/> VSAP1F VSAP1F	<input type="checkbox"/> VSETSF VSETSF VSHAML
<input type="checkbox"/> VSAP1F VSAP1F VSAP1F	<input type="checkbox"/> VSETSF VSGREF
<input type="checkbox"/> VSCEBP VSCEBP	<input type="checkbox"/> VSETSF VSHAML VSLEFF
<input type="checkbox"/> VSAP1F VSCEBP VSSP1F	<input type="checkbox"/> VSETSF VSPIT1
<input type="checkbox"/> VSAP1F VSGATA	<input type="checkbox"/> VSETSF VSRXRF
<input type="checkbox"/> VSAP1F VSGREF	<input type="checkbox"/> VSETSF VSSMAD VSSP1F
<input type="checkbox"/> VSAP1F VSHAML	<input type="checkbox"/> VSETSF VSSP1F

Compositionally Similar *Cis*-element Clusters in Coordinately Regulated Genes

12

Upper Limit on Number of Clusters: 2672

Matching Criteria:

Download All

Genes in Fetal_Liver_18

	Base Sequence	Second Sequence
<input type="radio"/> On All Selected		
<input checked="" type="radio"/> On All But Selected		
<input type="checkbox"/>	F13B human Coagulation factor XIII, B polypeptide	F13b mouse Coagulation factor XIII, beta subunit c
<input type="checkbox"/>	FGA human Fibrinogen alpha chain chr4	Fga mouse Fibrinogen, alpha polypeptide chr3
<input type="checkbox"/>	ANGPTL3 human Angiopoietin-like 3 chr1	Angptl3 mouse Angiopoietin-like 3 chr4
<input type="checkbox"/>	ACADSB human Acyl-Coenzyme A dehydrogenase, short/	Acadbs mouse Acyl-Coenzyme A dehydrogenase, short/
<input type="checkbox"/>	HAL human Histidine ammonia-lyase chr12	Hal mouse Histidine ammonia lyase chr10
<input type="checkbox"/>	COL2A1 human Collagen, type II, alpha 1 (primary o	Col2a1 mouse Procollagen, type II, alpha 1 chr15
<input type="checkbox"/>	FGB human Fibrinogen beta chain chr4	Fgb mouse Fibrinogen, B beta polypeptide chr3
<input type="checkbox"/>	PANK1 human Pantothenate kinase 1 chr10	Pank1 mouse Pantothenate kinase 1 chr19
<input type="checkbox"/>	IGF2 human Insulin-like growth factor 2 (somatomed	Igf2 mouse Insulin-like growth factor 2 chr7
<input type="checkbox"/>	ACSL4 human Acyl-CoA synthetase long-chain family	Acs14 mouse Acyl-CoA synthetase long-chain family
<input type="checkbox"/>	MTTP human Microsomal triglyceride transfer protei	Mtp mouse Microsomal triglyceride transfer protei
<input type="checkbox"/>	SLC2A2 human Solute carrier family 2 (facilitated	Slc2a2 mouse Solute carrier family 2 (facilitated
<input type="checkbox"/>	AFP human Alpha-fetoprotein chr4	Afp mouse Alpha fetoprotein chr5
<input type="checkbox"/>	OTC human Ornithine carbamoyltransferase chrX	Otc mouse Ornithine transcarbamylase chrX
<input type="checkbox"/>	TM4SF4 human Transmembrane 4 L six family member 4	Tm4sf4 mouse Transmembrane 4 superfamily member 4
<input type="checkbox"/>	C5 human Complement component 5 chr9	Hc mouse Hemolytic complement chr2
<input type="checkbox"/>	SERPINA7 human Serpin peptidase inhibitor, clade A	Serpina7 mouse Serine (or cysteine) peptidase inhi
<input type="checkbox"/>	ITIH2 human Inter-alpha (globulin) inhibitor H2 ch	Ith2 mouse Inter-alpha trypsin inhibitor, heavy c
Total = 18		Clear Gene Selection

5

OR

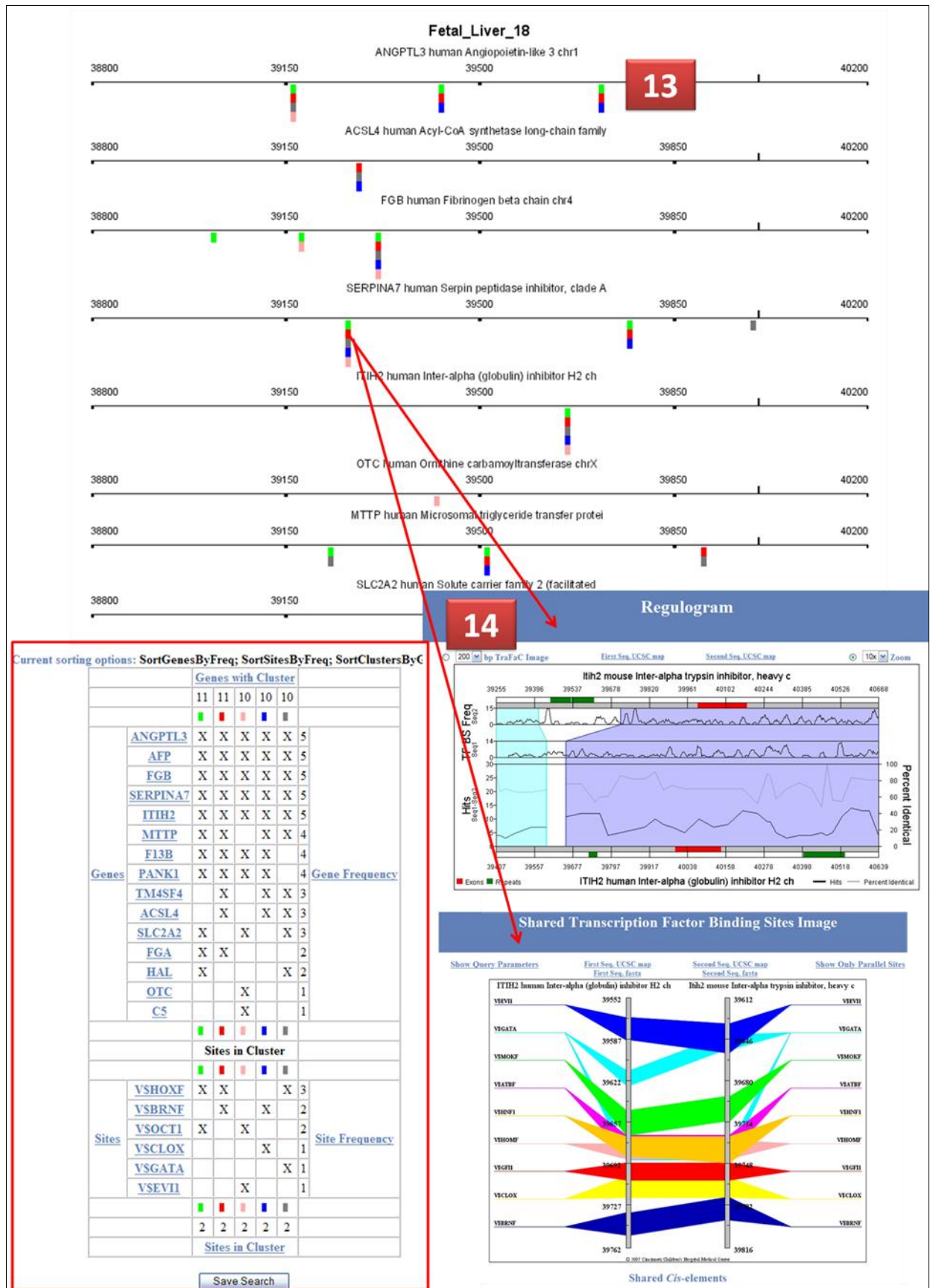
From :

To :

View Clusters

[Clear Cluster Selections](#)
[Select Clusters](#)

Select	Cluster ID	No. of Sites	No. of Ortholog Genes	Site Ids	Find Additional Target Genes
<input checked="" type="checkbox"/>	15534231	2	11	VSHOXF VSOCT1	ConCisE Scan
<input checked="" type="checkbox"/>	15532790	2	11	VSB RN F VSHOXF	ConCisE Scan
<input checked="" type="checkbox"/>	15533913	2	10	VSGATA VSHOXF	ConCisE Scan
<input checked="" type="checkbox"/>	15532126	2	10	VSB RN F VSCLOX	ConCisE Scan
<input checked="" type="checkbox"/>	15533704	2	10	VSEVT1 VSOCT1	ConCisE Scan

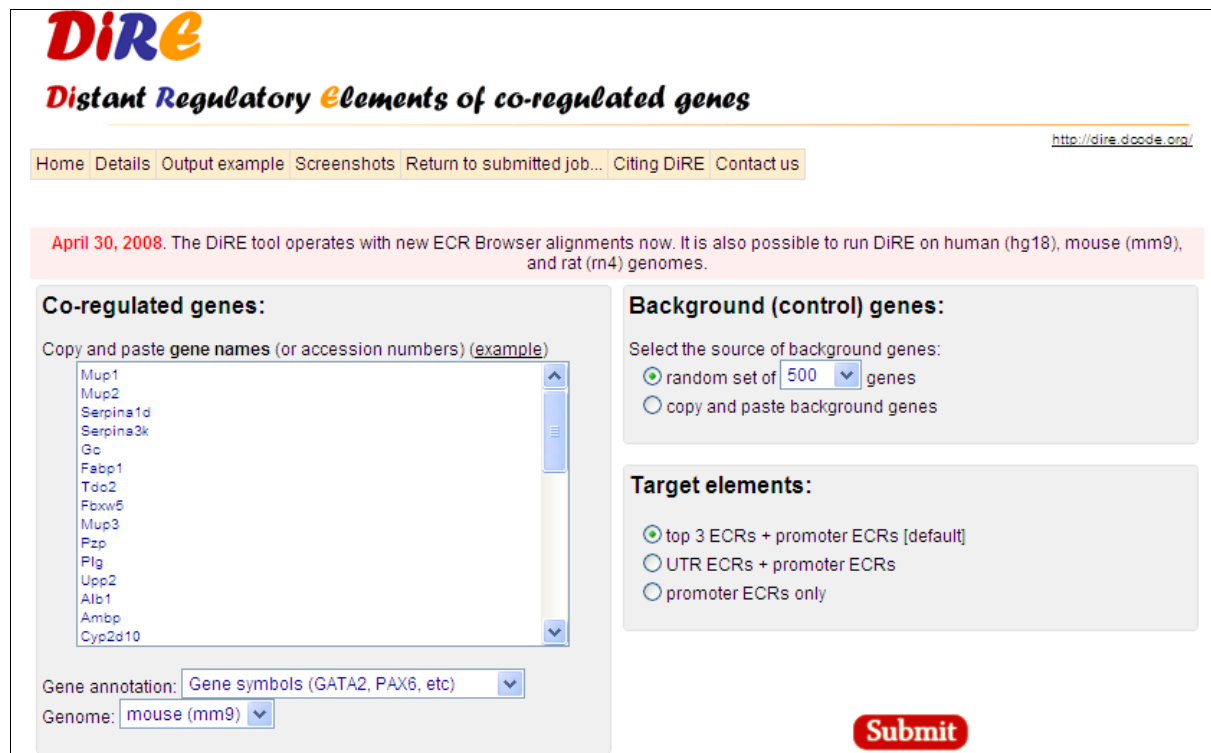


DiRE (Distant Regulatory Elements of co-regulated genes)**URL:** <http://dire.dcode.org>**Access:** Free; Web-based

Description/Utility: DiRE uses gene co-expression data, comparative genomics, and combinatorics of transcription factor binding sites (TFBSs) to find TFBS-association signatures that can be used for discriminating specific regulatory functions. DiRE's unique feature is the detection of REs outside of proximal promoter regions, as it takes advantage of the full gene locus to conduct the search. DiRE can predict common REs for any set of input genes for which the user has prior knowledge of co-expression, co-function, or other biologically meaningful grouping.

Input: Human or mouse or rat gene symbols or RefSeq accession numbers or chromosomal location.

Output: Graphical output and table.



DiRE
Distant Regulatory Elements of co-regulated genes

[Home](#) [Details](#) [Output example](#) [Screenshots](#) [Return to submitted job...](#) [Citing DiRE](#) [Contact us](#)

<http://dire.dcode.org/>

April 30, 2008. The DiRE tool operates with new ECR Browser alignments now. It is also possible to run DiRE on human (hg18), mouse (mm9), and rat (rn4) genomes.

Co-regulated genes:
Copy and paste **gene names** (or accession numbers) (example)

Mup1
Mup2
Serpina1d
Serpina3k
Gc
Fabp1
Tdo2
Fbxw5
Mup3
Pzp
Plg
Upp2
Alb1
Ambp
Cyp2d10

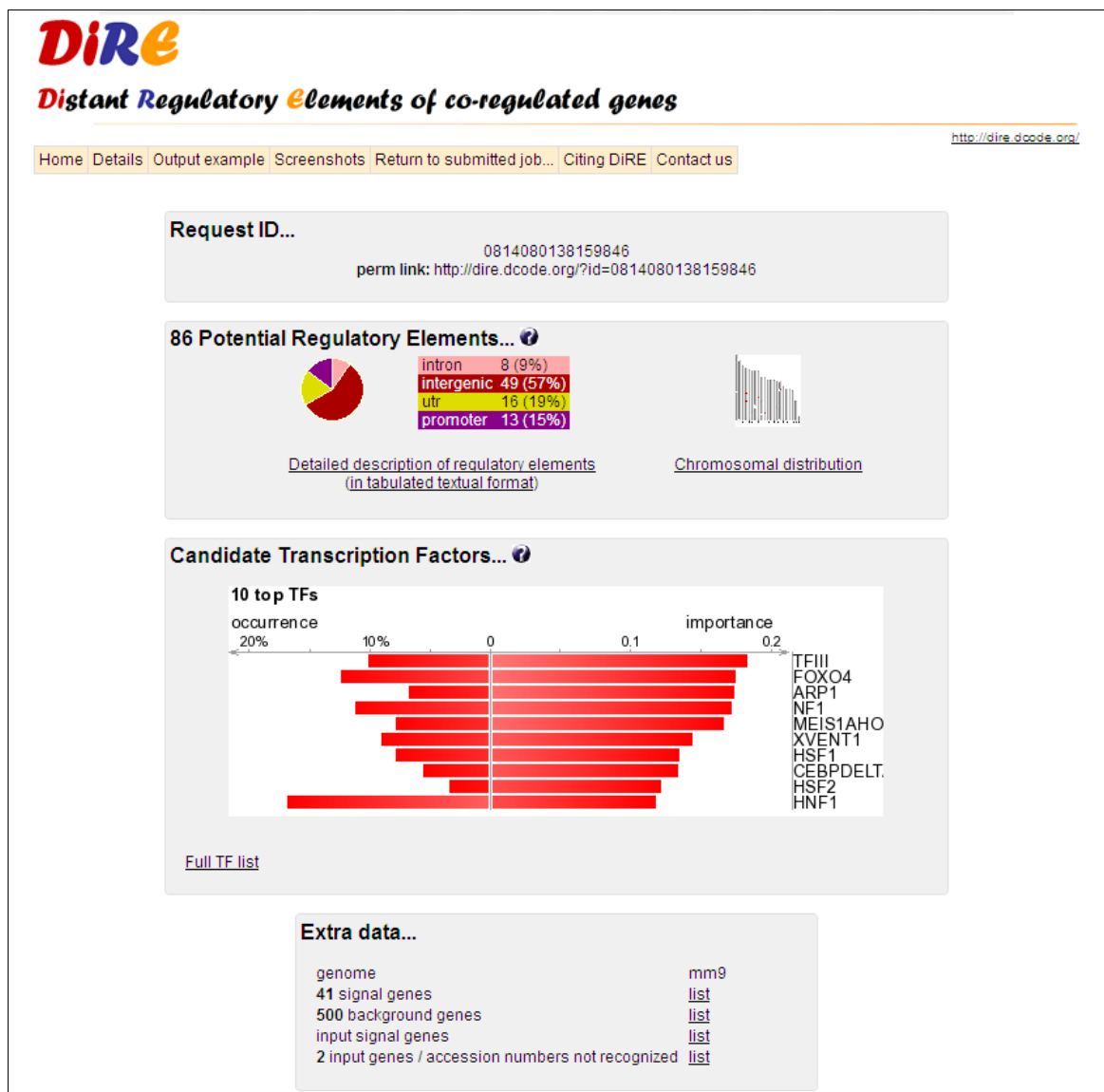
Gene annotation: **Gene symbols (GATA2, PAX6, etc)**

Genome: **mouse (mm9)**

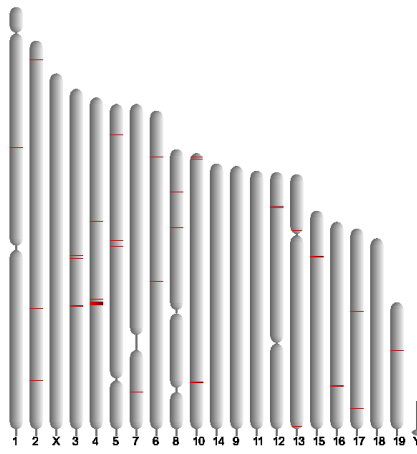
Background (control) genes:
Select the source of background genes:
☒ random set of **500** genes
☐ copy and paste background genes

Target elements:
☒ top 3 ECRs + promoter ECRs [default]
☐ UTR ECRs + promoter ECRs
☐ promoter ECRs only

Submit



1	chr1:132539023-132539510	intron	0.248	chr1:132531974-132585706	C4bp	4	::	AMEF2(90)	FOXJ2(134)	CRX(370)	DEAF1(435)
2	chr1:132585599-132586168	intergenic	1.009	chr1:132531974-132585706	C4bp	2	::	FOXJ2(92)	HNF1(98)		
3	chr10:24647246-24647516	UTR5	1.987	chr10:24633309-24888722	Arg1	5	::	EGR2(79)	TCF11(91)	TBX5(170)	CREB1(CJUN)(198)
4	chr10:24648558-24648656	promoter	0.491	chr10:24633309-24888722	Arg1	2	::	LPOLYA(21)	POU1F1(31)		
5	chr10:24711150-24711909	intergenic	0.248	chr10:24633309-24888722	Arg1	6	::	LHX3(34)	HLF(240)	FXR(242)	TBP(266)
6	chr10:24802988-24803646	intergenic	0.441	chr10:24633309-24888722	Arg1	10	::	KAISO(69)	NF1(151)	EFC(153)	CETS168(193)
7	chr10:127335414-127335700	intergenic	1.141	chr10:127316488-127335581	Rdh7	7	::	HNF4(87)	GCNF(90)	SMAD4(128)	ZTA(19)
8	chr10:128366645-128367417	intergenic	3.939	chr10:128360587-128395340	Itga7	11	::	TFII(191)	CMYB(267)	SRF(293)	RP(19)
9	chr10:128369507-128369715	promoter	0.264	chr10:128360587-128395340	Itga7	4	::	PAX4(52)	RSRFC4(98)	TBP(98)	CACCC(19)
10	chr10:128370121-128370435	promoter	0.389	chr10:128360587-128395340	Itga7	2	::	IRF(96)	CREB(237)		
11	chr12:104976179-104976530	intergenic	3.283	chr12:104976409-105087176	Serpina1d	7	::	HNF1(239)	DBP(274)	COUP(21)	
12	chr12:105011656-105011951	UTR5	0.140	chr12:104976409-105087176	Serpina1d	5	::	PAX6(97)	HNF1(196)	DBP(231)	HNF4(19)
13	chr12:105012145-105012287	promoter	0.170	chr12:104976409-105087176	Serpina1d	1	::	AP2ALPHA(46)			
14	chr12:105041910-105042366	intergenic	3.131	chr12:104976409-105087176	Serpina1d	6	::	XVENT1(93)	DEAF1(152)	PEA3(19)	
15	chr12:105503419-105503962	intergenic	3.772	chr12:105492760-105625335	Serpina3k	8	::	MZF1(89)	PPARA(118)	DEAF1	
16	chr12:105548838-105549321	intergenic	5.884	chr12:105492760-105625335	Serpina3k	15	::	DR4(27)	MEIS1(40)	TAL1(5)	
17	chr12:105596337-105596913	intergenic	1.865	chr12:105492760-105625335	Serpina3k	3	::	GCNF(185)	NRSE(320)	HSF1(19)	
18	chr13:4456075-4456256	intron	1.300	chr13:4283393-4499286	Akr1c6	2	::	COMP1(45)	PAX6(156)		
19	chr13:94331280-94331965	intergenic	4.066	chr13:94269679-94424505	Bhmt	16	::	MEF3(91)	ZTC1(97)	HFH8(105)	FOXO4(107)
20	chr15:82251872-82252090	intergenic	0.802	chr15:82224483-82282791	Cyp2d10	1	::	E2(143)			
21	chr15:82638016-82638715	intergenic	3.169	chr15:82640591-82640551	Cyp2d26	6	::	ARP1(243)	TBX5(247)	GCM(323)	APOLYA(469)
22	chr16:22896988-22897346	intergenic	1.449	chr16:22880451-22916411	Aheg	5	::	E2(10)	KAISO(147)	STAT(199)	HNF1(220)
23	chr16:22891821-22892187	UTR5	0.502	chr16:22880451-22916411	Aheg	3	::	TBP(267)	POU6F1(270)	PAX4(365)	
24	chr16:23093010-23093392	intergenic	4.806	chr16:23029218-23107816	Kng1	14	::	P53(70)	HSF1(122)	CETS168(123)	STAT(126)
25	chr16:23093434-23094542	intergenic	6.592	chr16:23029218-23107816	Kng1	40	::	HNF4(85)	COUP(98)	HNF4(99)	NF1(136)
26	chr17:12512398-12512765	intergenic	0.725	chr17:12511529-12612748	Plg	2	::	RFK(219)	BRN2(310)		
27	chr17:12536706-12537042	intergenic	0.441	chr17:12511529-12612748	Plg	7	::	FXR(96)	COUP(206)	NF1(226)	HOMA4(230)
28	chr17:12571303-12571533	UTR5	0.502	chr17:12511529-12612748	Plg	5	::	MZF1(5)	HNF1(125)	PAX4(127)	CREB(178)
29	chr17:57367501-57367733	UTR5	1.983	chr17:57333675-57372019	C3	9	::	NFKB(88)	HSF1(94)	CEBPB(99)	SREBP(152)
30	chr19:39348956-39349265	intergenic	2.794	chr19:39261363-39463746	Cyp2c29	5	::	GCM(78)	AMEF2(92)	TST1(95)	VDR(142)
31	chr19:39361390-39361543	promoter	1.342	chr19:39261363-39463746	Cyp2c29	6	::	COUP(36)	HNF4(37)	HNF4_DR1(37)	PPAR_DR1(37)



#	Transcription Factor	Occurrence	Importance
1	TFIIII	10.11%	0.18265
2	FOXO4	12.36%	0.17458
3	ARP1	6.74%	0.17360
4	NF1	11.24%	0.17135
5	MEIS1AHXA9	7.87%	0.16584
6	XVENT1	8.99%	0.14382
7	HSF1	7.87%	0.13469
8	CEBPDELTA	5.62%	0.13308
9	HSF2	3.37%	0.12143
10	HNF1	16.85%	0.11798
11	PAX8	10.11%	0.11440
12	MZF1	14.61%	0.11320
13	MAZR	14.61%	0.10547
14	CEBPB	12.36%	0.10042
15	GATA4	3.37%	0.09723
16	AHRARNT	7.87%	0.09635
17	HNF4_DR1	6.74%	0.09270
18	AP2ALPHA	14.61%	0.08329
19	CLOX	2.25%	0.08315
20	E2	6.74%	0.08006
21	PXR	5.62%	0.07557
22	TAL1	4.49%	0.07297
23	TEF1	6.74%	0.06742
24	XFD1	4.49%	0.06685
25	CACCCBINDINGFACTOR	6.74%	0.06362
26	HNF4	21.35%	0.06138
27	CETS168	6.74%	0.05554
28	ZTA	7.87%	0.05506
29	LPOLYA	11.24%	0.05337
30	ARNT	6.74%	0.05225
31	CREBP1CJUN	3.37%	0.05140

oPOSSUM (Distant Regulatory Elements of co-regulated genes)


URL: <http://burgundy.cmmt.ubc.ca/oPOSSUM/>

Access: Free; Web-based

Description/Utility: oPOSSUM is a system for determining the over-representation of transcription factor binding sites (TFBS) within a set of (co-expressed) genes as compared with a pre-compiled background set. The input is a set of gene identifiers. Analysis parameters are chosen. The system then compares the number of hits for each selected TFBS in the target gene set against the background set. Two different measures of statistical significance are applied to determine which sites are over-represented in the target set. The results of the analysis are displayed in tabular form.

Input: Human or mouse gene symbols or RefSeq accession numbers or Ensembl ID, or Entrez Gene IDs.

Output: Tab-delimited file.



oPOSSUM [About](#) | [Contact](#)

Welcome to oPOSSUM

oPOSSUM is a web-based system for the detection of over-represented transcription factor binding sites in the promoters of sets of genes.

Human SSA [Enter >>](#)

Human Single Site Analysis (SSA) is designed to detect over-represented conserved **single** sites in human and mouse genes.

Reference: Ho Sui, et al. (2005). oPOSSUM: Identification of over-represented transcription factor binding sites in co-expressed genes. *NAR*, 33(10):3154-64. PMID: 15933209

Human CSA (Module analysis) [Enter >>](#)

Human Combination Site Analysis (CSA) identifies over-represented **combinations** of conserved transcription factor binding sites in sets of human and mouse genes.

Reference: Huang, S., Fulton, D., et al. (2006). Identification of over-represented combinations of transcription factor binding sites in sets of co-expressed genes. *In Advances in Bioinformatics and Computational Biology*, Vol. 3. Imperial College Press, London, UK. 247-56. [E2F](#)

Worm SSA [Enter >>](#)

Worm Single Site Analysis (SSA) identifies over-represented conserved transcription factor binding sites in sets of *C. elegans* and *C. briggsae* genes.

Yeast SSA [Enter >>](#)

Yeast Single Site Analysis (SSA) identifies over-represented transcription factor binding sites in sets of *S. cerevisiae* genes. Phylogenetic footprinting has not been used for yeast.

Select Analysis Parameters

STEP 1: Enter a list of co-expressed genes

Species:☒ human ☐ mouse**Gene ID type:**☐ Ensembl ☒ HUGO/MGI Symbol/Alias ☐ RefSeq ☐ Entrez Gene☒ Paste gene IDs:

SLC38A4
SLC01B3
TM4SF4
UGT2B28
UGT2B4

☐ OR upload a file containing a list of gene identifiers:

STEP 3: Select parameters

Level of conservation:

 ▼

Matrix match threshold:

 ▼ %

Amount of upstream / downstream sequence:

 ▼

Number of results to display:

☒ Top ▼ results☐ OR only results with **Z-score** >= ▼ and **Fisher score** <= ▼ (Default values have been chosen based on empirical studies)

Sort results by:

☐ Z-score ☒ Fisher scorePress the **Submit** button to perform the analysis or **Reset** to reset the analysis parameters to their default values. Depending on server load

Analysis Results

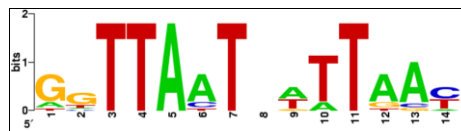
Selected Parameters

Conservation level: Top 10% of conserved regions (min. conservation 70%)
Matrix match score: 80%
Upstream sequence length: 2000
Downstream sequence length: 2000
Number of genes submitted: 26
Number of genes included: 21
Number of genes excluded: 5

Target Genes

Analyzed: OTC ACSL4 NR1H4 AFP HAL APOB C5 SERPINA7 ANGPTL3 MTTP SLC38A4 F13B COL2A1 ITIH2 PANK1 IGF2 SLC2A2 FGA FGB TM4SF4 ACADSB
Excluded: ADH6 CYP3A7 SLC01B3 UGT2B28 UGT2B4

Gene ID	Ensembl ID	Chr	Strand	TSS	Promoter Start	Promoter End	TFBS Sequence	TFBS Start	TFBS Rel. Start	TFBS End	TFBS Rel. End	TFBS Orientation	TFBS Score
OTC	ENSG00000018477	X	+	38094822	38094822	38094820	38097939	1538	38097947	1544	-1	0.892	
				38096821	38094821	38096820	38097939	1119	38097947	1127	-1	0.892	
				38096821	38094821	38096820	38096611	1791	38096619	1799	1	0.806	
ACSL4	ENSG00000069266	X	-1	108793289	108791290	108795289	108794700	-1411	108794708	-1419	1	0.973	
				108863277	108861276	108865277	108861830	1448	108861836	1440	1	0.809	
				108863277	108861276	108865277	108864545	-1268	108864553	-1276	1	0.801	
				108863277	108861276	108865277	108864561	-1284	108864569	-1292	-1	0.861	
				108863277	108861276	108865277	108864870	-1293	108864878	-1301	-1	0.825	
				108863277	108861276	108865277	108864872	-1195	108864880	-1163	1	0.808	
NR1H4	ENSG00000012304	12	+	89421269	89419269	89423268	89421120	-149	89421128	-141	-1	0.824	
				89421269	89419269	89423268	89421188	-81	89421186	-73	1	0.849	
				89421269	89419269	89423268	89423943	1679	89423951	1683	1	0.810	
AFP	ENSG00000081051	4	+	74520797	74520797	74522796	74520377	-420	74520385	-412	1	0.840	
				74520797	74520797	74522796	74520648	-149	74520656	-141	1	0.846	
				74526887	74524887	74528886	74526790	-97	74526798	-89	1	0.846	
HAL	ENSG00000084110	12	-1	84914202	84912203	84916202	84914224	-122	84914232	-130	-1	0.929	



Unknown/Novel

Non-conserved

MEME – Multiple Em for Motif Elicitation

URL: <http://meme.sdsc.edu/meme/meme.html>

Access: Free; Web-based; The web-based version has limit of number of base pairs (should not be more than 60,000 bp). Alternately, you can download the software and install it on your own computer. This will allow you to use many features that are not available with the interactive version.

Description/Utility: MEME is a tool for discovering motifs in a group of related DNA or protein sequences. A motif is defined as a sequence pattern that occurs repeatedly in a group of related protein or DNA sequences. MEME represents motifs as position-dependent letter-probability matrices which describe the probability of each possible letter at each position in the pattern. Individual MEME motifs do not contain gaps. Patterns with variable-length gaps are split by MEME into two or more separate motifs.

MEME takes as input a group of DNA or protein sequences (the training set) and outputs as many motifs as requested. MEME uses statistical modeling techniques to automatically choose the best width, number of occurrences, and description for each motif.

MEME sends you three e-mail messages:

- a confirmation message,

- the MEME results, and
- the MAST results of searching the training set for the motifs found by MEME using MAST.

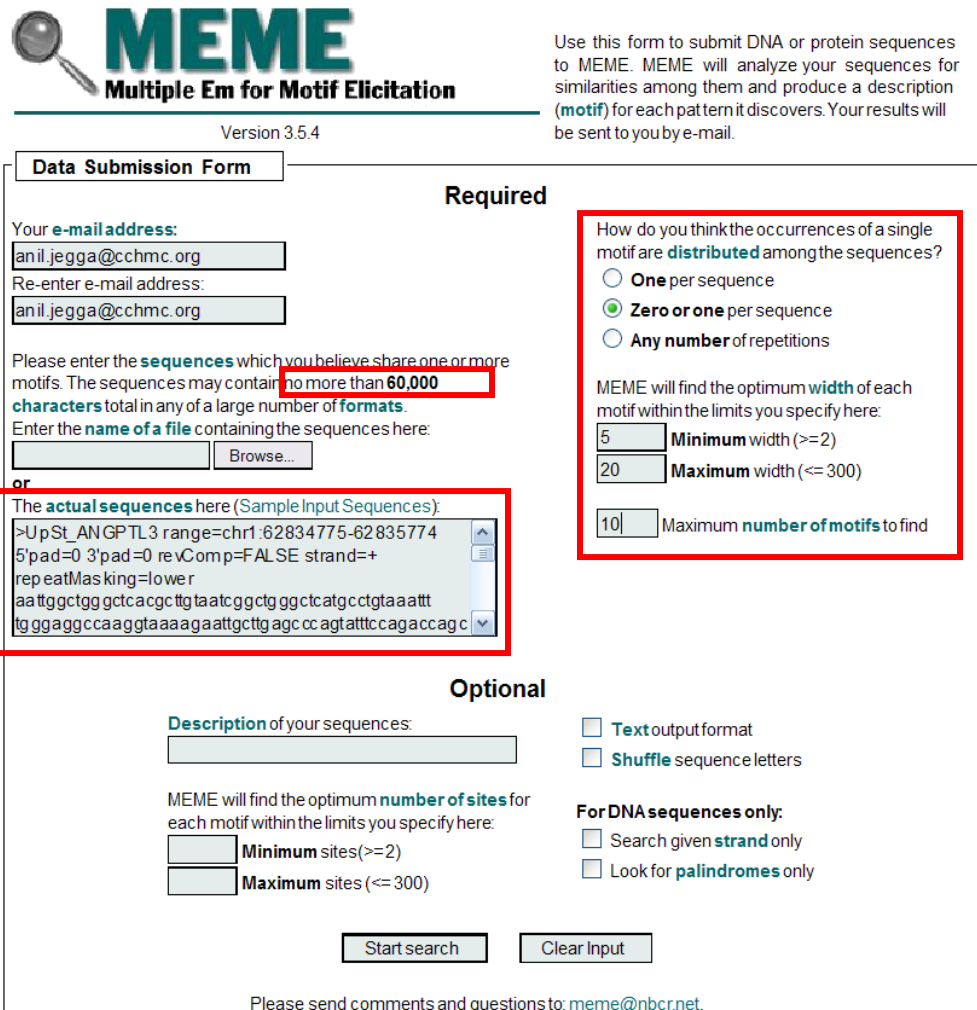
One of the features of MEME that come handy is it facilitates you to compare the identified motifs to known TFBSs. This will help you to find out potentially “real” novel motifs in the sequences of coexpressed genes.

Input: Multi-fasta files (not more than 60,000 bp)

Output: There is no good way of using/storing the graphical output (especially if you want to use it in a presentation or publication) except for taking a screen capture.

Drawbacks/Limitations:

- Larger sequences can take considerable amount of time (more than a day).
Explore the mirror sites OR download the application and use it locally.
- Results can be difficult to interpret.
- Always try to mask the repeat elements otherwise your top scored motifs could be repeat elements!



Use this form to submit DNA or protein sequences to MEME. MEME will analyze your sequences for similarities among them and produce a description (**motif**) for each pattern it discovers. Your results will be sent to you by e-mail.

Version 3.5.4

Data Submission Form

Required

Your **e-mail address**:

 Re-enter e-mail address:

Please enter the **sequences** which you believe share one or more motifs. The sequences may contain **no more than 60,000 characters** total in any of a large number of **formats**.
 Enter the **name of a file** containing the sequences here:

or

The **actual sequences** here (Sample Input Sequences):

How do you think the occurrences of a single motif are **distributed** among the sequences?
☐ One per sequence
☒ Zero or one per sequence
☐ Any number of repetitions

MEME will find the optimum **width** of each motif within the limits you specify here:
 Minimum width (>=2)
 Maximum width (<= 300)
 Maximum number of motifs to find

Optional

Description of your sequences:

MEME will find the optimum **number of sites** for each motif within the limits you specify here:
 Minimum sites (>=2)
 Maximum sites (<= 300)

☐ **Text** output format
☐ **Shuffle** sequence letters

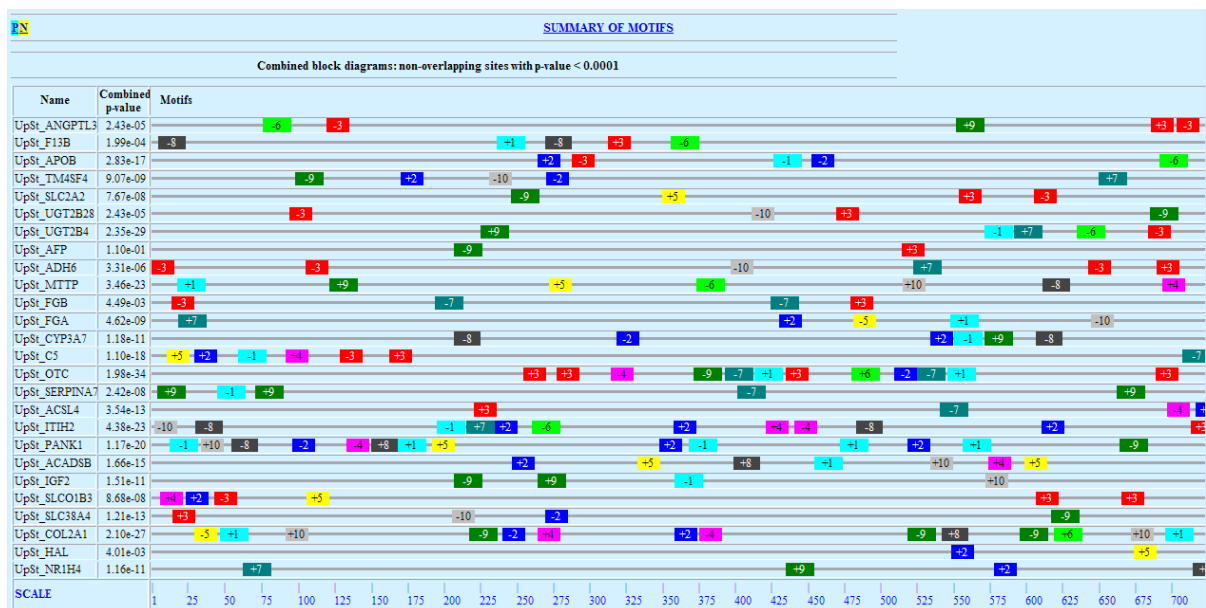
For DNA sequences only:
☐ Search given **strand** only
☐ Look for **palindromes** only

Please send comments and questions to: meme@nbcrl.net.

Your MEME search results will be sent to: anil.jegga@cchmc.org
If you do not receive a confirming email message, there could be an error in your email address.

- E-mail address: anil.jegga@cchmc.org
- Sequence file: **pasted_sequences**
- Description:
- Distribution of motif occurrences: **Zero or one per sequence**
- Number of different motifs: **10**
- Minimum number of sites:
- Maximum number of sites: **33**
- Minimum motif width: **5**
- Maximum motif width: **20**
- Statistics on your dataset:

type of sequence	dna
number of sequences	33
shortest sequence (residues)	1000
longest sequence (residues)	1000
average sequence length (residues)	1000.0
total dataset size (residues)	33000



Weeder

Similar to MEME but comparative studies have shown it be better than MEME (speed, output, results, robustness)

Drawbacks/Limitations: Results are sent in the mail message. Some email clients (e.g. Groupwise mail) truncate longer messages and therefore you may not see complete results (especially if you are using several sequences). I use Gmail and that works fine.

MOA TOOL

Home

Online tools

MOA & Weeder

RNA motifs

Additional tools

MOA browser

MOA profile calculator

Downloads

MOA 1.0.1

RNA motifs 1.0

MOA TOOL

Tools for MOA Discovery in nucleotide sequences

Weeder

Motif discovery in sequences from co-regulated genes

Version 1.0.1 running

Click here to switch to WeederH

Please note: submitting simultaneously a large (> 10) number of jobs has the effect of slowing down the server, for your jobs, as well as the jobs submitted by other users. If you plan to use Weeder extensively, you can download the stand-alone version. Client IPs and e-mail addresses generating high workloads on the server (as defined in the previous sentence) might have their jobs terminated before completion without notice.

Enter your e-mail address:

Input at least two sequences

Input sequences (FASTA)

```
gattgtatgactaatgtctgcctatgacacagccgaagagaga
tgctgtctgacagacagagccgcctgtatgacacacatcagg
```

from: Homo sapiens

To upload a file, first locate it by using the browse button, then click on Upload.

☒ Check here if you want to look for motifs in both strands of the input sequences

☐ Check here if you want motifs to appear in all the sequences (default is some)

Hint: don't try this option even if you're pretty much sure that all your sequences share a motif.

☐ Check here if you think that the motif might appear more than once in a single sequence (without, you expect zero or one occurrence per sequence)

And, finally, you'd like:

☐ a quick scan (short motifs, no longer than 8 nt) of your sequences

☒ a normal scan of your sequences

☐ a complete and thorough scan

Important: input larger than 200K will be limited to quick analysis. For large jobs, you can download the source code by following the link in the bottom page.

Quick scan: results will be ready in a few minutes. Normal scan: results will be ready in one-two hours. Thorough scan: results will be ready in a few hours. However, the normal scan is fast. If nothing interesting comes out, try the thorough one.

Name of this job:

Click submit once to start the computation. Click reset to clear all the fields.

Weeder

Thank you!

You submitted 33 sequences from Homo sapiens

You asked for a normal scan

You asked to process both strands of the input sequences

You asked for a normal scan

A confirmation e-mail and the final results will be sent to the following e-mail address:

and.jegga@gmail.com

*** Your Weeder Web Results ***

The name of this job was Fetal_Liver_33_27

Input sequences from H. sapiens

You asked to include both strands of the input sequences

You asked for a normal scan of your sequences

Confused about this output? Click [here](#)

Searching for motifs of length 6 with 1 mutations....

- CGATTGA 0.81
- TAAAGC 0.70
- ATTGAT 0.67
- TATGAT 0.63
- GATTGA 0.61
- ATGGTA 0.60
- TCATTG 0.59
- TGGTAT 0.59
- TGATTA 0.59
- TGATAT 0.58

Searching for motifs of length 8 with 2 mutations....

- CGTTTGA 0.93
- ACTAAGC 0.88
- GATAAGC 0.87
- TATGAT 0.87
- CTAAGC 0.87
- AGTATTC 0.84
- ACATTGAT 0.82
- GTAATACT 0.80
- CTAGCAAT 0.79
- ATATGTC 0.78

*** Interesting motifs (highest-ranking) seem to be ***

GATAAAT
AGTTTATC

0 redundant motifs found:

Best occurrences (match percentage):

Seq 1: GATAAAT 205 (92.24)

1 + [AATAAAT] 676 (85.29)

1 + [ATTAATC] 922 (88.60)

1 - [ATAAAT] 786 (92.79)

1 - [AATAAAT] 697 (92.36)

1 - [ATAAAT] 169 (85.17)

2 + [TAAATAAT] 508 (85.63)

2 + [TATAAAT] 944 (85.73)

2 - [AAAAAT] 956 (85.28)

2 - [ATAAAT] 776 (85.29)

2 - [ATAAAT] 652 (90.33)

4 + [AATAAAT] 546 (87.13)

4 + [AATAAAT] 756 (85.24)

5 + [AATAAAT] 393 (85.29)

6 + [TATAAAT] 260 (85.24)

7 - [ATAAAT] 430 (94.77)

8 + [AATAAAT] 307 (87.13)

8 + [AATAAAT] 791 (87.13)

8 - [AAAAAAT] 808 (85.19)

8 - [AATAAAT] 484 (85.29)

8 - [TATAAAT] 285 (85.24)

8 - [TATAAAT] 13 (87.66)

9 + [ATAAAT] 603 (100.00)

9 + [AATAAAT] 615 (85.24)

9 - [ATAAAT] 438 (94.77)

10 + [ATAAAT] 603 (100.00)

10 + [AATAAAT] 615 (85.24)

11 + [AATAAAT] 438 (94.77)

11 + [ATAAAT] 148 (85.10)

11 - [AATAAAT] 205 (85.77)

12 + [AATAAAT] 143 (92.93)

12 - [ATAAAT] 271 (92.79)

12 + [ATAAAT] 286 (87.13)

12 + [ATAAAT] 523 (90.60)

12 - [ATAAAT] 896 (94.77)

12 - [ATAAAT] 347 (85.29)

13 + [ATAAAT] 549 (92.36)

13 - [ATAAAT] 832 (88.34)

13 - [ATAAAT] 577 (88.34)

13 - [ATAAAT] 484 (85.29)

14 - [ATAAAT] 832 (88.34)

14 - [ATAAAT] 577 (88.34)

16 + [ATAAAT] 143 (92.93)

16 + [ATAAAT] 316 (89.17)

16 + [ATAAAT] 814 (85.29)

16 - [ATAAAT] 967 (92.36)

16 - [ATAAAT] 940 (90.40)

18 - [ATAAAT] 637 (89.17)

18 - [ATAAAT] 515 (85.29)

Frequency Matrix

	All Occs	Best Occs
A	28	16
C	16	17
G	16	17
T	16	26
A	0	0
C	0	0
G	0	25
T	0	0
A	208	6
C	9	19
G	26	0
T	25	0
A	198	10
C	13	21
G	25	0
T	0	0
A	7	16
C	25	28
G	1	0
T	0	25

Conserved

Currently, there are no publicly available tools that can look for conserved motifs across multiple sequences. You can use MEME or Weeder by using orthologous promoters. But this can sometimes lead to erroneous results (for e.g. the high scoring motifs can be the same ones just coming from ortholog copy of the same gene).

In case anyone needs assistance in this type of analysis (for e.g. a genome-wide scan for two or multispecies conserved motifs), we can assist (the tool/application we are working on is in development stage).

	GTGGGTGCCCT	TGGCTTCTGGAAAT	CCGGCTT
Human	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Chimp	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Rhesus	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Bushbaby	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
TreeShrew	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Mouse	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
GuineaPig	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Dog	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Cat	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Horse	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Cow	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Armadillo	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Elephant	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Tenrec	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Opossum	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg
Platypus	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg	ccgccgtgggtgccctcgtggcggttcttggaatgcgccattctg

Chapter 3: Functional Enrichment Analysis of the Transcriptome

Objectives

1. What is the functional enrichment for a given set of genes
 - a. Gene Ontology (Biological Process, Molecular Function and Cellular Component)
 - b. Pathways (Kegg, Biocarta, etc.)
 - c. Protein Domains (Interpro, PFAM, etc.)
 - d. Functional Keywords (SwissProt keywords – genes associated with controlled vocabulary terms e.g. all genes associated with the word “apoptosis”)
2. What is the expression pattern of these genes in other conditions (or how do my genes score in other microarray expression experiments)
3. How to prioritize/rank the genes in my gene list so that I can select a handful for further experimental validation

Introduction

The pathway, ontology data sources and analysis tools establish a basis for finding links between lists of genes in their associated biological network context. Several tools/servers are available that effectively utilize these various resources and help in understanding of the unifying biological themes underlying one's data. Here I list some of the “popular” and useful ones. **My favorites** are:

1. For single gene list enrichment analysis: DAVID, MSigDB, FatiGO+ and Panther (not necessarily in that order)
2. For comparison:
 - a. Two gene lists: FatiGO+ (extensive parameter coverage including miRNAs!)
 - b. Multiple gene lists: Panther (but the parameters are limited)
3. For comparison of a gene list with other published gene sets based on different experiments: L2L and MSigDB (for cancer related datasets Oncomine is probably the best)
4. For both enrichment analysis and also candidate gene prioritization: ToppGene. It's probably the only one database currently that takes into account literature co-citations and also mouse phenotype data for functional enrichment analysis.

Tools and Servers

DAVID

Description: Database for Annotation, Visualization and Integrated Discovery (DAVID) provides a comprehensive set of functional annotation tools for investigators to understand biological meaning behind large list of genes.

URL: <http://david.abcc.ncifcrf.gov/>

DAVID Bioinformatics Resources 2007
National Institute of Allergy and Infectious Diseases (NIAID), NIH

Home | Start Analysis | Shortcut to DAVID Tools | Technical Center | Downloads & APIs | Term of Service | Why DAVID? | About Us

Welcome to DAVID Bioinformatics Resources 2003 - 2007

The Database for Annotation, Visualization and Integrated Discovery (DAVID) 2007 is the fifth program of DAVID 2006, a comprehensive set of functional annotation tools to understand biological meaning from a gene list, DAVID tools are:

- Identify enriched biological themes, particularly GO terms
- Discover enriched functional-related gene groups
- Cluster redundant annotation terms
- Visualize genes on BioCarta & KEGG pathway maps
- Display related many-genes-to-many-terms on 2-D view.
- Search for other functionally related genes not in the list
- List interacting proteins
- Explore gene names in batch
- Link gene-disease associations
- Highlight protein functional domains and motifs
- Redirect to related literatures
- Convert gene identifiers from one type to another.
- And more

Screen Shot 1

Please cite the web site or [Genome Biology 2003; 4\(3\):P3](#) within any publication that makes use of any methods inspired by DAVID.

 SAC
 SAC Frederick
 NIAID
 ABCC
 NCI Frederick
 NCI
 FIRSTGOV
 Department of Health and Human Services

[Term of Service](#) | [Contact Us](#) | [Site Map](#)

Annotation Summary Results

Current Gene List: demolist1 171 DAVID IDs
Current Background: Homo sapiens Check Defaults [X] Clear All

☐ Main Accessions (0 selected)
☐ Other Accessions (0 selected)
☒ Gene Ontology (3 selected)

Term	Count	Percentage	Chart
GOTERM_BP_1	136	79%	[Chart]
GOTERM_BP_2	131	76%	[Chart]
GOTERM_BP_3	127	74%	[Chart]
GOTERM_BP_4	119	69%	[Chart]
GOTERM_BP_5	104	60%	[Chart]
GOTERM_BP_ALL	136	79%	[Chart]
GOTERM_CC_1	121	70%	[Chart]
GOTERM_CC_2	106	61%	[Chart]
GOTERM_CC_3	95	55%	[Chart]
GOTERM_CC_4	86	50%	[Chart]
GOTERM_CC_5	65	38%	[Chart]
GOTERM_CC_ALL	121	70%	[Chart]
GOTERM_MF_1	129	75%	[Chart]
GOTERM_MF_2	119	69%	[Chart]
GOTERM_MF_3	103	60%	[Chart]
GOTERM_MF_4	97	56%	[Chart]
GOTERM_MF_5	78	45%	[Chart]
GOTERM_MF_ALL	129	75%	[Chart]

☒ Protein Domains (3 selected)
☒ Pathways (3 selected)
☐ General Annotations (0 selected)
☐ Functional Categories (3 selected)
☐ Protein Interactions (0 selected)
☐ Literature (0 selected)
☒ Disease (1 selected)

Term	Count	Percentage
GENETIC_ASSOCIATION_DB	23	13%
OMIM_DISEASE	32	18%

Combined View for Selected Annotation
Functional Annotation Clustering

2D View

corresponding gene-term association positively reported
corresponding gene-term association not reported yet

Options: Run Using Options

epidermy, progressive myoclonus type 2a, lafora disease [dfe...
 protein phosphatase 1, regulatory subunit 1d
 glutamate decarboxylase 2 [neuronal, soluble, and brain, GAD-65...
 spectrin, beta, erythrocytic [includes spherocytosis, clinical ty...
 tumor protein p53 [B-fragment, cyclin...
 lectin, galactose-binding, soluble, 3 [galactin 3]
 p61 and bin domain 1
 immunoglobulin heavy constant gamma 1 (IgM marker)
 immunoglobulin heavy locus
 immunoglobulin kappa variable 1d-13
 v-jb-10 erythroid leukemia viral oncogene homolog 2, neuro[glial]blastoma deriv...
 3-hydroxyanthranilate 3,4-dioxygen...
 arachidonate 15-lipoxygenase
 myeloperoxidase
 cytochrome p450, family 4, subfamily a, polypeptid...
 cytochrome p450, family 3, subfamily a, polypeptid...
 hemoglobin, beta
 hemoglobin, delta
 hemoglobin, alpha 1

MSigDB (Molecular Signatures Database)

Description: The “annotation” features helps you to compute overlaps between your gene set and other gene sets in MSigDB. Additionally, you can also categorize members of the gene set by gene families and display the gene set expression profile based on a selected compendium of expression profiles. The analysis results include:

Statistics:

- overlaps shown lists the number of overlapping gene sets displayed in the report: By default, the report displays the 10 gene sets in the collection that best overlap with your gene set. If you compute overlaps from the Annotations page, you can choose the number of overlapping gene sets to display in the report.
- gene sets in collection lists the total number of gene sets being analyzed
- genes in comparison lists the number of genes in your gene set
- genes in collection lists the number of unique genes in the gene sets being analyzed

Descriptions of the overlapping gene sets, including

- Link to the gene set card
- Number of genes in the gene set
- Description of the gene set
- Number of genes in the overlap between this gene set and your gene set
- *p value* indicating the significance of the overlap
- Color bar shading from light green to black, where **lighter colors indicate more significant p values (< 0.05) and black indicates less significant p values (≥ 0.05).**


Overlap matrix showing the genes in the overlapping gene sets

- Rows list the genes in your gene set, with gene descriptions and links to gene annotations
- Columns list the overlapping gene sets, with links to the gene set cards
- Overlaps are computed using HUGO gene symbols. In rare instances, a gene set may contain a gene symbol that is not in the GENE_SYMBOL chip annotation file. Such gene symbols are ignored when overlaps are computed and appear crossed out in the matrix. If a gene set has a source platform other than GENE_SYMBOL, each gene symbol in the gene set is translated to its probe identifier(s) on the source platform. The matrix lists the probe identifier(s) in parentheses following the gene symbol. If a gene symbol cannot be translated, a question mark (?) appears in place of the probe identifier(s).

URL: <http://www.broad.mit.edu/gsea/msigdb/annotate.jsp>

Access: Free for academics (need to register)

Others: Explore the GSEA (Gene Set Enrichment Analysis) and also Gene Pattern


MSigDB
Molecular Signatures Database

[Home](#)
[Software](#)
[MSigDB](#)
[Docs](#)
[Resources](#)

Name: Region
Input: Region
Output: None Selected
Mode: Image Capture

Annotations

Explore gene set annotations to gain further insight into the biology behind a gene set in question:

- compute overlaps with other gene sets in MSigDB ([details](#))
- categorize members of the gene set by gene families ([details](#))
- display the gene set expression profile based on a selected compendium of expression profiles ([details](#))

MSigDB
[MSigDB home](#)
[collections](#)
[search](#)
[annotations](#)
[gene sets](#)
[gene families](#)
[help](#)

Genes to annotate
ABCC2
ALDH1B1
ARF1
ATP6V0E1
BHMT
CDH1
CDX1
CLDN7
CNN2
CPT1A
DGCR6
DGCR6L
EIF1AP1
FBXW4
FKBP4

Compute overlaps with
☐ C1: Chromosomal locations
☒ C2: Curated gene sets
☐ CP: Canonical pathways
☐ CGP: Chemical and genetic perturbations
☒ C3: Motif gene sets
☒ TFT: Transcription factor targets
☒ MIR: miRNA targets
☒ C4: Computed gene sets
overlaps:
show genesets ☒ clustered

Compendia expression profiles
☒ Human tissue compendium (Novartis)
☐ Global Cancer Map (Broad Institute)
☐ NCI-60 cell lines (National Cancer Institute)

Gene families

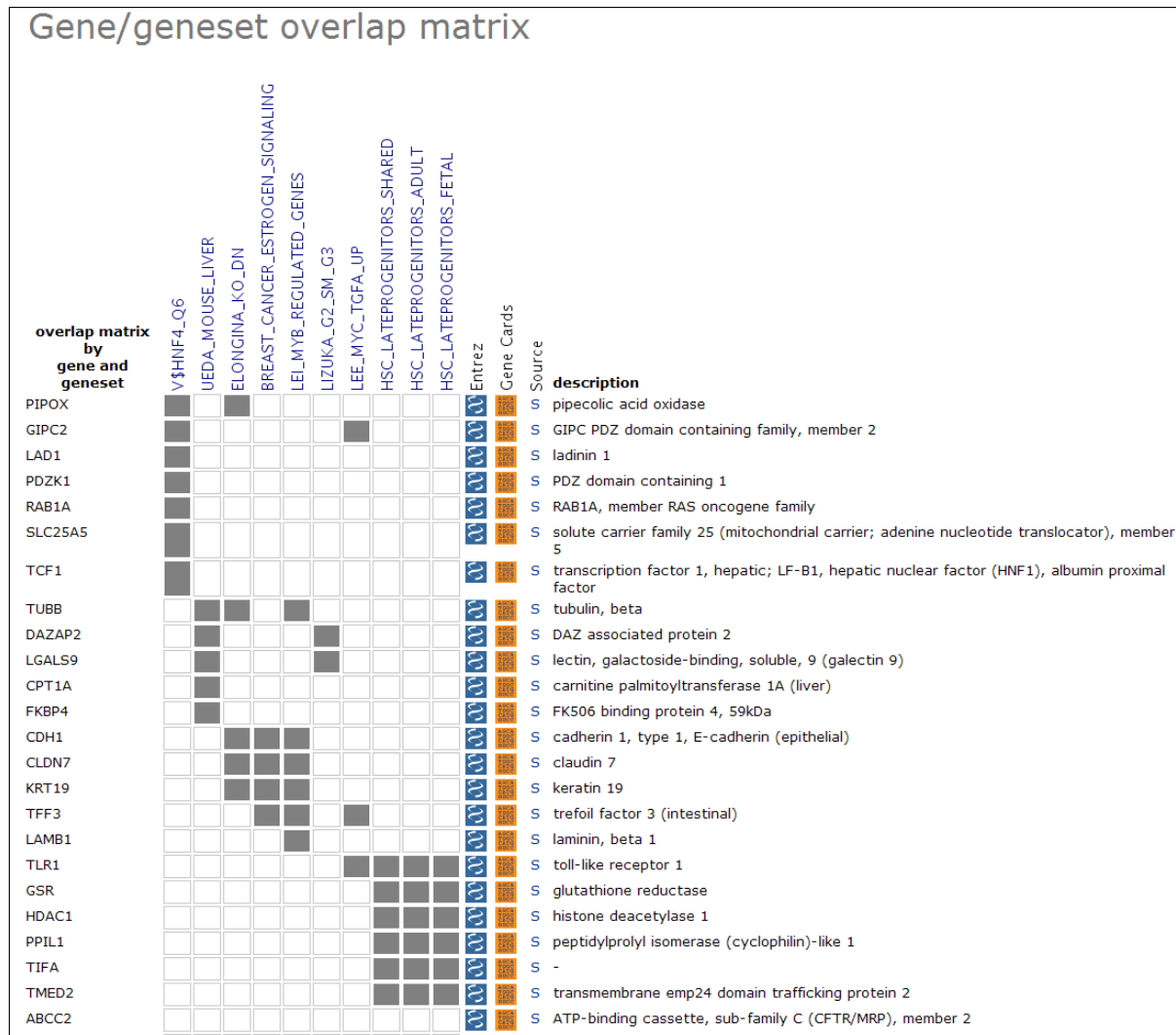
Chip format
GENE SYMBOL

Overlap results

collections	# overlaps shown	# genesets in collections	# genes in comparison (n)	# genes in collections (N)
C2, CP, CGP, C3, TFT, MIR	10	2524	55	21826

Click the gene set name to see the gene set card. Click the number of genes [in brackets] to download the list of genes.

Geneset name [# genes (K)]	description	# genes in overlap (k)	k/K	p value ⁽²⁾
V\$HNF4_Q6 [273]	Genes with promoter regions [-2kb,2kb] around transcription start site containing the motif AARGT...	7	<div style="width: 3.2%;"></div>	4.99 e ⁻⁶
UEDA_MOUSE_LIVER [165]	Genes identified as time indicators in mouse liver.	5	<div style="width: 2.4%;"></div>	5.59 e ⁻⁵
ELONGINA_KO_DN [184]	Downregulated in MES cells from elongin-A knockout mice	5	<div style="width: 2.3%;"></div>	9.29 e ⁻⁵
BREAST_CANCER_ESTROGEN_SIGNALING [101]	Genes preferentially expressed in breast cancers, especially those involved in estrogen-receptor...	4	<div style="width: 1.8%;"></div>	1.17 e ⁻⁴
LEI_MYB_REGULATED_GENES [325]	Myb-regulated genes	6	<div style="width: 2.8%;"></div>	1.47 e ⁻⁴
LIZUKA_G2_SM_G3 [9]	Genes highly expressed in poorly differentiated vs. moderately differentiated hepatocellular carc...	2	<div style="width: 0.9%;"></div>	2.21 e ⁻⁴
LEE_MYC_TGFA_UP [61]	Genes up-regulated in hepatoma tissue of Myc+Tgfa transgenic mice	3	<div style="width: 1.4%;"></div>	4.74 e ⁻⁴
HSC_LATEPROGENITORS_SHARED [463]	Up-regulated in mouse hematopoietic late progenitors from both adult bone marrow and fetal liver ...	6	<div style="width: 2.8%;"></div>	9.06 e ⁻⁴
HSC_LATEPROGENITORS_ADULT [470]	Up-regulated in mouse hematopoietic late progenitors from adult bone marrow (Late Progenitors Sha...	6	<div style="width: 2.8%;"></div>	9.76 e ⁻⁴
HSC_LATEPROGENITORS_FETAL [473]	Up-regulated in mouse hematopoietic late progenitors from fetal liver (Late Progenitors Shared + ...	6	<div style="width: 2.8%;"></div>	1.01 e ⁻³

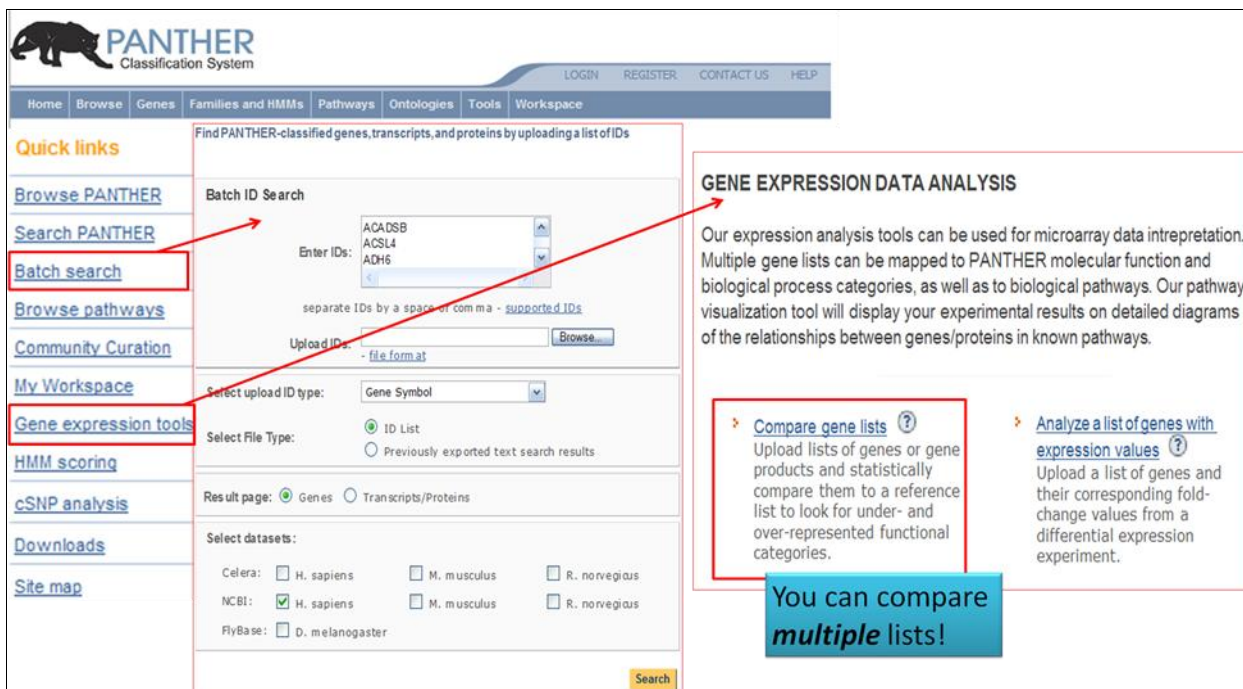


Panther

Description: The PANTHER (Protein ANalysis THrough Evolutionary Relationships) Classification System is a unique resource that classifies genes by their functions, using published scientific experimental evidence and evolutionary relationships to predict function even in the absence of direct experimental evidence. Proteins are classified by expert biologists into families and subfamilies of shared function, which are then categorized by molecular function and biological process ontology terms. For an increasing number of proteins, detailed biochemical interactions in canonical pathways are captured and can be viewed interactively.

URL: <http://www.pantherdb.org>

Access: Free web-based. By registering, you can store you gene lists and results (“workspace”).



PANTHER
Classification System

Home Browse Genes Families and HMMs Pathways Ontologies Tools Workspace

Quick links

Browse PANTHER

Search PANTHER

Batch search

Browse pathways

Community Curation

My Workspace

Gene expression tools

HMM scoring

cSNP analysis

Downloads

Site map

Find PANTHER-classified genes, transcripts, and proteins by uploading a list of IDs

Batch ID Search

Enter IDs: ACADSB
ACSL4
ADH6

separate IDs by a space or comma - supported IDs

Upload IDs:

Select upload ID type: Gene Symbol

Select File Type: ☒ ID List ☐ Previously exported text search results

Result page: ☒ Genes ☐ Transcripts/Proteins

Select datasets:

Celera: ☐ H. sapiens ☐ M. musculus ☐ R. norvegicus

NCBI: ☒ H. sapiens ☐ M. musculus ☐ R. norvegicus

FlyBase: ☐ D. melanogaster

GENE EXPRESSION DATA ANALYSIS

Our expression analysis tools can be used for microarray data interpretation. Multiple gene lists can be mapped to PANTHER molecular function and biological process categories, as well as to biological pathways. Our pathway visualization tool will display your experimental results on detailed diagrams of the relationships between genes/proteins in known pathways.

Compare gene lists ?
Upload lists of genes or gene products and statistically compare them to a reference list to look for under- and over-represented functional categories.

Analyze a list of genes with expression values ?
Upload a list of genes and their corresponding fold-change values from a differential expression experiment.

You can compare multiple lists!

Select lists to analyze

For example, you can upload a two lists, one of up-regulated genes and one of down-regulated genes, from a differential mRNA microarray experiment.

UPLOAD OR SELECT LIST FROM YOUR WORKSPACE

Upload list:

List type: ? Please select list type...

Upload list:

If there are redundant IDs, only the first will be used in the analysis.

Choose list from your workspace

☐ Root Folder

Uploaded and selected lists:

☒ FetalLiverSpecific.txt

☒ FetalBrainSpecific.txt

Compare Classifications of Lists ?

Map lists of genes to a PANTHER ontology. For pathways, you can then view the gene expression values overlaid on top of a pathway diagram, where genes will be colored differently for different clusters of genes.

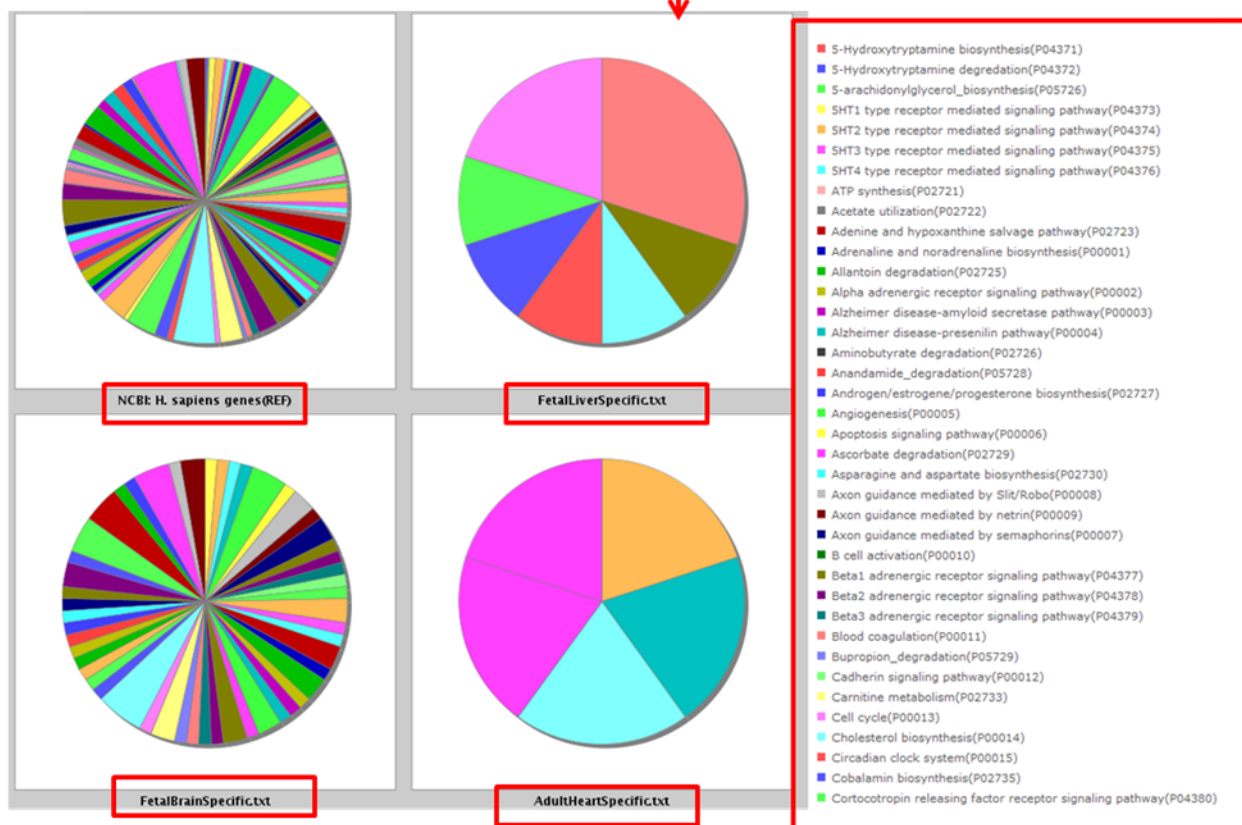
Use the binomial statistics tool to compare classifications of multiple clusters of lists to a reference list to statistically determine over- or under- representation of PANTHER classification categories. Each list is compared to the reference list using the binomial test ([Cho & Campbell, TIGs 2000](#)) for each molecular function, biological process, or pathway term in PANTHER.

Steps:
1. Select list(s) to analyze
2. Select reference list

1. Select Lists to Compare to a Reference List
For example, each selected list may be a cluster of co-expressed genes under a particular set of conditions.
 selected: FetalLiverSpecific.txt
FetalBrainSpecific.txt
AdultHeartSpecific.txt

2. Select Reference List
For example, the reference list may be the set of all genes in the experiment, or the set of all genes in the genome being analyzed.
 default: NCBI: H. sapiens genes

Search options
PANTHER Ontology:
☒ Pathways
☐ Biological Process
☐ Molecular Function
☒ Use the Bonferroni correction for multiple testing ?



L2L

Description: L2L is a database of published microarray gene expression data, and a software tool for comparing the published data to a user's own microarray results.

URL: <http://depts.washington.edu/l2l/>

ToppGene

Description: The majority of common diseases are multi-factorial and modified by genetically and mechanistically complex polygenic interactions and environmental factors. High-throughput genome-wide studies like linkage analysis and gene expression profiling, tend to be most useful for classification and characterization but do not provide sufficient information to identify or prioritize specific disease causal genes. Hypothesizing that the majority of genes that impact or cause disease share membership in any of several functional relationships ToppGene integrates several data sources for disease candidate gene prioritization. ToppGene for the first times uses mouse phenotype data as one of the features for gene prioritization and we have observed that using mouse phenotype data greatly improves the human disease candidate gene analysis and prioritization

URL: <http://toppgene.cchmc.org>

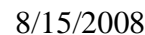
Access: Free, web-based

Utility: Can be used for both functional enrichment in gene lists and also to prioritize candidate genes.

Input: Human gene symbols or gene IDs (NCBI's Entrez gene IDs)

Output: Graphical and results are downloadable as tab-delimited text files.

Limitations: Currently works only for human genes.



Babelomics (FatioGO)

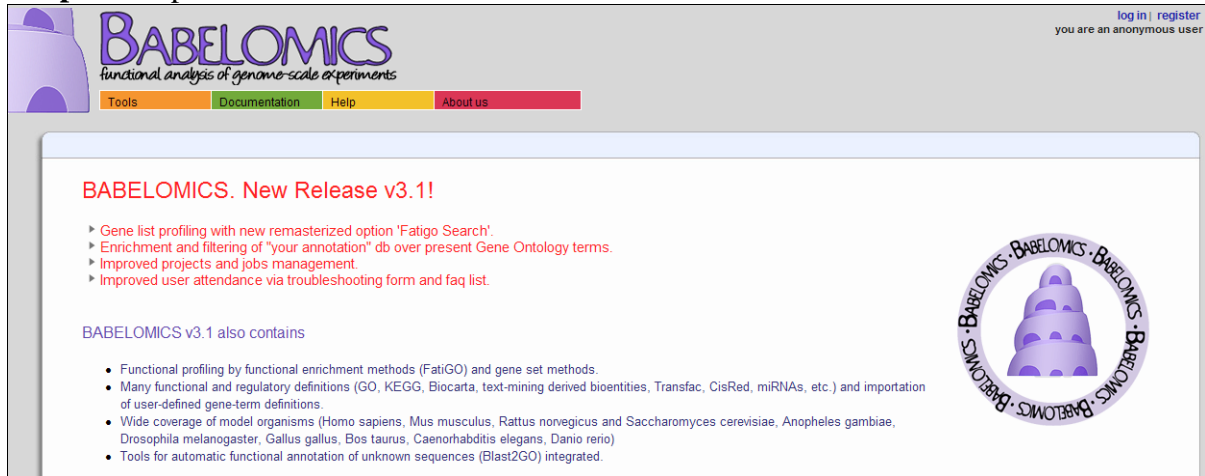
Description: In addition to GO terms it can test simultaneously for KEGG pathways, Interpro motifs, SwissProt keywords, TFBSs and CisRed motifs. The distribution of any combination (or all) of the terms between two groups of genes can be simultaneously tested by means of a Fisher exact test. All the P-values are adjusted by FDR.

URL: <http://www.fatigo.org>

Access: Free web-based

Input: Gene symbols

Output: Graphical and downloadable as text files



The screenshot shows the Babelomics website homepage. At the top, there is a navigation bar with links for Tools, Documentation, Help, and About us. The main content area features a red banner for "BABELOMICS. New Release v3.1!". Below this, there are several bullet points highlighting new features: Gene list profiling with 'Fatio Search', enrichment and filtering of annotations, improved projects and jobs management, and improved user attendance via troubleshooting and FAQ. A circular logo with the word "BABELOMICS" repeated around a central graphic is also visible. The footer mentions "BABELOMICS v3.1 also contains" followed by a list of features like functional profiling, various gene sets, model organisms, and automatic functional annotation.



The screenshot shows the FatioGO web interface for functional enrichment. The title is "FatioGO Functional enrichment". A descriptive paragraph explains that FatioGO performs functional enrichment analysis by comparing two lists of genes using Fisher's exact test, with various gene modules and criteria supported. The interface includes tabs for "Compare", "Your annotations", "Genomics", and "Search". Below these, there is a section for "Organism" with a dropdown menu. The main part of the interface is divided into two sections: "List of Genes #1" and "List of Genes #2". Each section has a text input for "A list of genes" and a file upload section for "or a gene file" with options for "file from your computer" (with a "Browse..." button) and "or from your projects". At the bottom, there is a checkbox for "Rest of genome".

FatiScan
 Gene set enrichment

FatiScan implements a segmentation test which checks for asymmetrical distributions of biological labels associated to genes ranked in a list (Al-Shahrour et al., 2005a,b). Unique in this type of approaches, this test only needs the list of ordered genes and not the original data which generated the sorting. This means that can be applied to the study of the relationship of biological labels to any type of experiment whose outcome is an sorted list of genes. Since Babelomics is linked to GEPAS, genes sorted by differential expression between two experimental conditions can be studied, but also genes correlated to a clinical variable (such as the level of a metabolite) or even to survival. Moreover, other lists of genes ranked by any other experimental or theoretical criteria can be studied (e.g. genes arranged by physico-chemical properties, mutability, structural parameters, etc.) in order to understand whether there is some biological feature (among the labels used) which is related to the experimental parameter studied.

FatiScan Your Annotations

Organism

List of Genes #1
 A list of genes

or a gene file

Databases

GO - biological process	<input type="checkbox"/>	options»
GO - molecular function	<input type="checkbox"/>	options»
GO - cellular component	<input type="checkbox"/>	options»
KEGG pathways	<input type="checkbox"/>	options»
Interpro motifs	<input type="checkbox"/>	options»
Swissprot keywords	<input type="checkbox"/>	options»
MicroRNA	<input type="checkbox"/>	options»
Transcription factors	<input type="checkbox"/>	options»
BioCarta	<input type="checkbox"/>	options»
cisRED	<input type="checkbox"/>	options»

Statistics
 Fisher exact test

Number of partitions

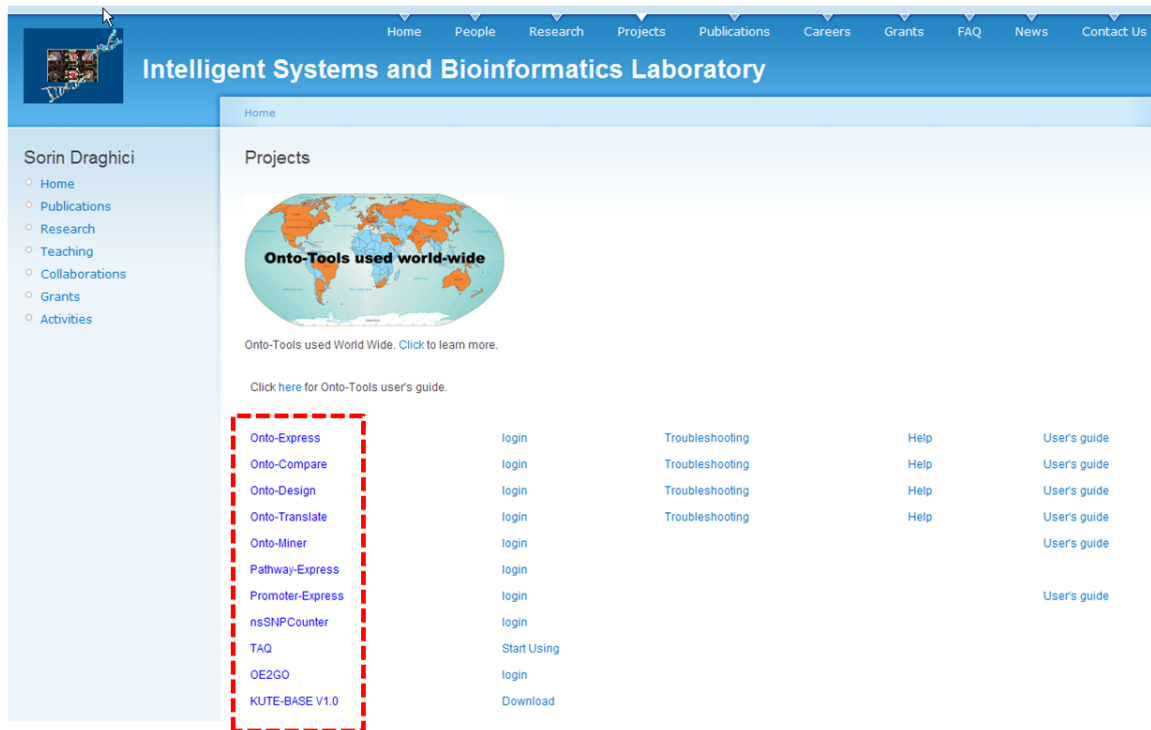
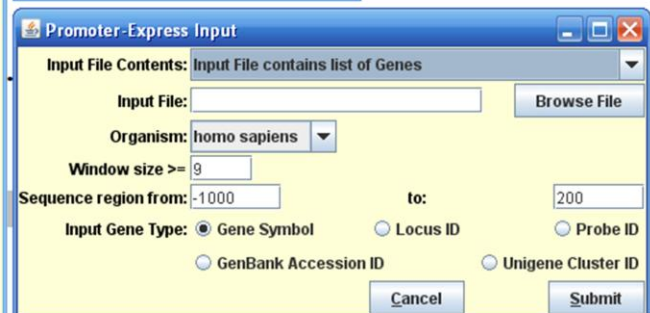
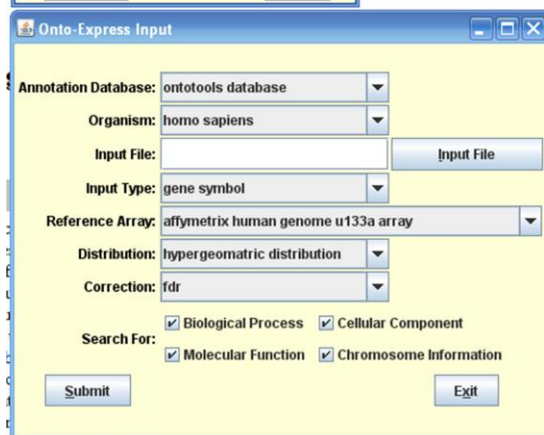
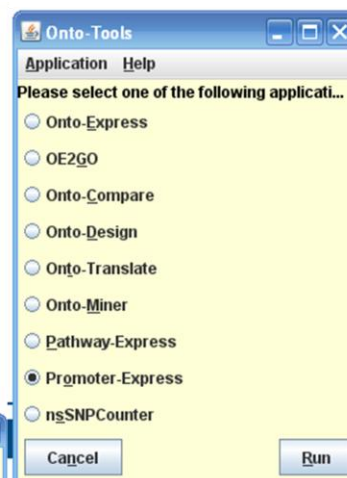
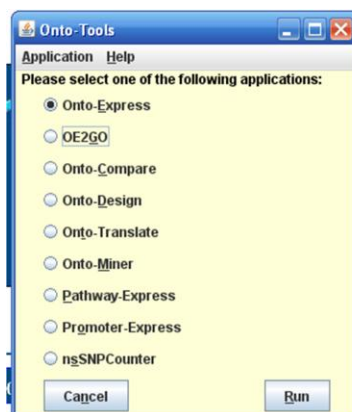
Sort genes/values?
☒ Greater to smaller
☐ Smaller to greater

OntoExpress

Description: OntoExpress constructs functional profiles (using Gene Ontology terms) for the following categories: biochemical function, biological process, cellular role, cellular component, molecular function and chromosome location. Statistical significance values are calculated for each category (Draghici et.al, Genomics, 81(2), 2003).

URL: <http://vortex.cs.wayne.edu/projects.htm>

<http://vortex.cs.wayne.edu/projects.htm>

Onto-Express

[Troubleshooting](#)
[Help](#)

The typical result of a microarray experiment is a list of tens or hundreds of genes found to be differentially regulated in the condition under study. Independently of the methods used to select these genes, the common task faced by any researcher is to translate these lists of genes into a better understanding of the biological phenomena involved. Currently, this is done through a tedious combination of searches through the literature and a number of public databases. We developed Onto-Express (OE) as a novel tool able to automatically translate such lists of differentially regulated genes into functional profiles characterizing the impact of the condition studied. OE constructs functional profiles (using Gene Ontology terms) for the following categories: biochemical function, biological process, cellular role, cellular component, molecular function and chromosome location. Statistical significance values are calculated for each category. We demonstrated the validity and the utility of this comprehensive global analysis of gene function by analyzing two breast cancer data sets from two separate laboratories. OE was able to identify correctly all biological processes postulated by the original authors, as well as discover novel relevant mechanisms (Draghici et al, Genomics, 81(2), 2003). Other results obtained with Onto-Express can be found in Ostermeier et al, Lancet, 360(9335), 2002.

Onto-Compare

[Troubleshooting](#)
[Help](#)

Microarrays are at the center of a revolution in biotechnology, allowing researchers to screen tens of thousands of genes simultaneously. Typically, they have been used in exploratory research to help formulate hypotheses. In most cases, this phase is followed by a more focused, hypothesis driven stage in which certain specific biological processes and pathways are thought to be involved. Since a single biological process can still involve hundreds of genes, microarrays are still the preferred approach as proven by the availability of focused arrays from several manufacturers. Since focused arrays from different manufacturers use different sets of genes, each array will represent any given regulatory pathway to a different extent. We argue that a functional analysis of the arrays available should be the most important criterion used in the array selection. We developed Onto-Compare as a database that can provide this functionality, based on the GO nomenclature.

Onto-Design

[Troubleshooting](#)
[Help](#)

Many Laboratories chose to design and print their own microarrays. At present, the choice of the genes to include on a certain microarray is a very laborious process requiring a high level of expertise. Onto-Design database is able to assist the designers of custom microarrays by providing the means to select genes based on their biological process, molecular function or cellular component.

Onto-Translate

[Troubleshooting](#)
[Help](#)

In the annotation world a same piece of information can be stored and viewed differently across different databases. For instance, more than one Affymetrix probe ids can refer to the same GenBank sequence (accession number) and more than one nucleotide sequence from GenBank can be grouped in a single UniGene cluster. The result of Onto-Express depends on whether the input list contains Affymetrix probe IDs, GenBank accession numbers or UniGene cluster IDs. The user has to be aware of relations between the different forms of the data in order to interpret correctly the results. Even if the user is aware of the relationships and knows how to convert them, most existing tools allow conversions of individual genes. Onto-Translate is a tool that allows the user to perform easily such translations.

Onto-Miner

Onto-Miner (OM) provide a single and convenient interface that allow the user to interrogate our databases regarding annotations of known genes. OM will return all known information about a given list of genes. Advantages of OM include the fact it allows queries with multiple genes and allows for scripting. This is unlike GenBank which uses a single gene navigation process.

Pathway-Express

The automated functional profiling approach of OE helps the researchers to better understand the biological phenomenon under study by pointing out statistically significant cellular functions. However, graphical representations of gene interactions (pathways) can be very useful. As more data becomes available, the question "is there a known pathway containing my gene(s) of interest?" will gradually transform into "how do I find the most interesting pathway(s) involving my gene(s)?"

Pathway-Express (PE) is a new tool in the Onto-Tools ensemble that is designed to answer such questions. Our goal is to provide a system that will automatically find such interesting pathways. When the user submits a list of genes, the system performs a search and builds a list of all associated pathways.

Chapter 4: Identification of Regulatory Regions: Using Trafac and Other Related Tools

*Before going any further, please check the **GenomeTrafac** database (<http://genometrafac.cchmc.org>) which has more than 12,000 human-mouse gene pairs and about 200 microRNAs already analysed for potential regulatory regions. The genes or microRNAs you are interested in might be already there. This will save you the trouble of uploading them through trafac. You DON'T need any account to access the database.*

Introduction

Trafac (<http://trafac.cchmc.org>) is a web-accessible system, developed by us at Biomedical Informatics, to detect and visualize constitutionally similar clusters of transcription factor binding sites between a pair of genes.

Method

We first identify regions of genomic sequence conservation between two related but yet divergent species. Potential transcription factor binding sites, based on the TRANSFAC Professional Library, are independently predicted for both the genomic sequences. A JAVA servlet is then used to parse the results of this sequence conservation and transcription factor binding sites (*cis*-elements) data into an Oracle database. The database is mined for the detection of clusters of *cis*-elements in common between the two genes.

Output

1. **Trafacgram:** A high-resolution graphical image depicting the relative structural arrangement of the shared transcription factor (TF) binding sites within each sequence.
2. **Regulogram:** To better understand the occurrence of conserved *cis*-element clusters as a function of entire genomic regions, rather than discrete homologous blocks, we developed a *cis*-element hit-density graph (Regulogram) that depicts the density of shared *cis*-elements occurring within a moving window through conserved regions.

Utility

1. To find conserved TF binding sites between two orthologous genes in the context of sequence similarity.
2. It can be a valuable filtration tool for identifying potential novel regulatory regions, hitherto unknown.
3. It helps in comparison of heterologous genes (for e.g. two genes with similar expression).
4. It helps in identifying shared TF binding sites within genes that exhibit coordinate expression.
5. Finally, it also helps in understanding the constitution of regulatory regions of tissue specific genes.

What do you need?

1. Genomic Sequences: Exons, Introns, upstream and downstream regions.
2. Exon coordinates or positions in the genomic sequence.
3. Masking the repeats.
4. List of transcription factor binding sites.
5. Sequence alignment data where applicable.

How to Use Trafac?

From the home page of Trafac you can approach to the analysis by taking either of the two routes.

1. Cis-element Clusters within BlastZ Alignments: To Find conserved cis-clusters within BLASTZ-identified conserved sequence alignment blocks. Using this link you would be able to visualize the alignment between an orthologous pair of genes (mostly human and mouse sequences). Most importantly, you can view the common putative TF binding sites shared by the human and mouse genes in the context of the conserved regions. This utility is limited to a pairwise comparison of only those sequences for which the alignment data is present in the database. But, then if you are interested in a particular gene(s) you can upload the requisite input data to visualize the results. Alternatively, you can send us the sequences and we will do the rest for you.
2. Cis-elements Shared Between any Gene Pair: To Find cis-element clusters between user-selected gene segment pairs. This link would take you to explore the genes for regulatory elements irrespective of the sequence similarity. The main advantage of this route is you can compare any gene with any other gene or known promoters/enhancers in the Trafac database. If you are analyzing a group of co-expressed or coordinately regulated genes, this approach is recommended, especially when you know the transcription start site.

You can search the trafac database either by the

- Accession number: A GenBank or Celera accession number can be used.
- Name/Symbol of the gene: Trafac supports the gene nomenclature approved by the HUGO.
- Description/any term: Enter any term, for example, human, mouse, repair, etc. Please make sure to enter only one term.

Sequence Group: You can also select the genes based upon the gene group. For example, selecting "DNA repair" from the list of the available groups would display all genes belonging to DNA repair group and which have the BlastZ alignment data entered to Trafac. *Please note that the list of genes under each of the groups at present is not exhaustive. We are in the process of building up the database and adding more genes and more groups. If you are interested in any particular gene or group please let us know so that we can add them to the Trafac database.*

Sequence Selector: Sequence selector page has two parts. The top one is the query part wherein you can enter one or more than one search criteria. The second part or the lower half displays your search results once you click on the search.

Using Sequence Selector for identifying potential conserved regulatory regions:

1. Enter one or more than one search criteria and click the Search button. Alternatively you can choose one from the existing groups of genes.
2. The results will be displayed in the lower half.
3. For example, using the term "Human" for description displays a list of entries in the lower half.
4. The results table has check boxes in the first column against each entry. You can check one or more to view the sequences.
5. Check your selected entries and click on Select. This would take you to the BlastZ alignments page. The first two columns of the table show the sequence information for the human and mouse sequences followed by a date of entry column. The last column shows three options. The **view** option would take you to the Local alignments page. This is nothing but a summary view of the sequence alignment information. The second option "**PIP**" leads to a graphic display of the alignment image. This PIP is generated using the PipMaker software. This is a pdf file so you need to have an Adobe Acrobat Reader installed to view this. The third option is the "**Regulogram**", a cis-element hit density graph in the context of sequence similarity.

Using Sequence Selector for identifying constitutionally similar regulatory sites:

1. Enter one or more than one search criteria and click the Search button. Alternatively you can choose one from the existing groups of genes
2. The results will be displayed in the lower half.
3. For example, using the term "Human" for description displays a list of entries in the lower half.
4. The results table has check boxes in the first column against each entry. You can check the first sequence and click. You will be prompted to select the second sequence. After selecting both the sequences, you will be led to the TraFaC query Page.
5. The **TraFaC Query Page** allows the user to alter the various parameters like sequence extent, matrices, image size, comparison parameters, whether based on matrix families or individual matrices, etc.

Uploading Sequences:

If you wish to upload the genes of your interest and those which are not already present in TraFac database, you need to have an account. You can obtain one by sending a mail (anil.jegga@chmcc.org or bhuvana.sakthivel@cchmc.org). If you have only one or two genes to be analyzed, you can mail us the sequences or GenBank accession number(s) or gene symbols and we can upload the genes for you.

Multi-Upload Page

From the Multi-Upload page, you can parse all the input files to the TraFac server and see the results. There are certain rules you need to follow when uploading the input files.

- All the input files should be strictly in the recommended format (in text format for all except the MatInspector files and the PIP, which are in html format and pdf respectively).
- An accession number should always be entered whenever you are uploading any of the files except MatInspector files.
- The exon file, repeat mask and the actual sequence file upload is optional. However, we advise you to upload all of these so that you can comprehend your results more clearly.

The requisite input files can be uploaded from your local system using the browse button. Click the upload button when you are through. Uploading may take some time depending upon your sequence size. You can upload any of these data in parts but do remember to associate with a unique ID, which is the accession number. An "upload successful" message indicates that you are ready to view your results.

Input Files

- a. Sequence Files
- b. Exon Files
- c. RepeatMasker Output Files
- d. PipMaker Output Files
- e. MatInspector/Match Output Files

1. **Sequence Files:** The nucleotide sequence files need to be in the fasta format.
2. **RepeatMasker:** The RepeatMasker web application is used to mask the repeats before aligning them. Given below are the brief instructions for using the RepeatMasker. You can find a detailed set on the RepeatMasker page. Use the 'browse' button to select the fasta file created above, or copy the fasta sequence into the text box. Select the "html" return format, the appropriate DNA Source and press Submit Sequence. Save the Matches as a text file to your computer by selecting File->Save As... from the main menu. You can use copy and paste to save the Repeat Mask output. The mask output alone is required. Do not save the masked sequence.
3. **Exon Files:** An optional text file, providing the positions of transcriptional units in the first sequence. The directionality of a gene (< or >), its start and end positions, and name should be on one line, followed by separate lines specifying the start-positions and end-positions of each exon. An optional line beginning with a "+" character can indicate the first and last nucleotides of the translated region (including the initiation codon, Met, and the stop codon). Blank lines are ignored. Exons must be specified in order of increasing address even if the gene is on the reverse strand (<). For example, the Exons file might begin as follows:
> 100 800 Gene 1
100 200
300 400
600 800
4. **PipMaker Output Files:** PipMaker computes alignments of similar regions in two DNA sequences. The resulting alignments are summarized with a Percent Identity Plot (PIP). It generates graphical output as a PDF document by default. For TraFaC,

you need the **Blastz alignment file**, the **summary file** and an optional **PIP** file. These are referred as "**text**", "**concise**" and "**pip**" files respectively in the PipMaker output data, which you receive by an E-mail. We give here the brief instructions for using the Advanced PipMaker. For detailed instructions refer the advanced PipMaker instructions page. Follow the Advanced PipMaker instructions (the files you have created above can be used). Enter your E-mail address. Press the Submit button. When the alignment is finished, you will receive an E-mail containing multiple attachments. Of the different output files **Trafac** needs only three of them *viz.*, **Concise Alignment file**: This file has a summary of the sequence alignment information. **Text file**: This is a verbose file and is actually a BLASTZ alignment file. **PIP**: a .pdf file of the percent identity plot. Store the first two files as text files.

5. **MatInspector Professional/Match Output Files**: You can use either of these programs for detection of transcription factor binding sites. MatInspector professional or Match is a tool that utilizes a library of matrix descriptions for transcription factor binding sites to locate matches in sequences of unlimited length. To access both these programs you need to have registered with them and it is free. You can find the detailed instructions on these pages. We however, would like to highlight some of the points like: Be certain to choose the appropriate matrix family e.g. vertebrates. And follow the default options for the rest of the parameters. When you have the output, save the MatInspector files as html files. But in case of Match files, save them as text files.

Once you have all of the requisite files, log on to Trafac. From Advanced Tools, select Upload/Parse Sequence Data.

Related Tools

Concise Scanner (Conserved Cis-Element Scanner; <http://concise-scanner.cchmc.org>):

Concise Scanner was developed primarily as an answer to the question as to what are the potential additional target genes for an identified regulatory module(s). Tools based on the phylogenetic footprinting like TraFaC (and GenomeTraFaC, the human-mouse gene regulatory region repository created using TraFaC server), and others while helping in identification of potential regulatory regions provide little or no information as to through what genes the transcription factors (TFs) exert their function in the living system. Providing a complement to the above listed phylogenetic approaches, we developed Concise (Conserved Cis Element) Scanner that undertakes a more targeted search, finding phylogenetically conserved regulatory targets of defined transcription factors whose DNA binding site specificity is known. It identifies potential targets of one or more clusters of transcription factors with a defined cis-regulatory target specificity, using human and mouse genomes. It enables you to select one or more transcription binding sites and search all genes in the GenomeTrafac database for clusters containing the selected site(s). Within each cluster, you can view the exact position of each binding site.

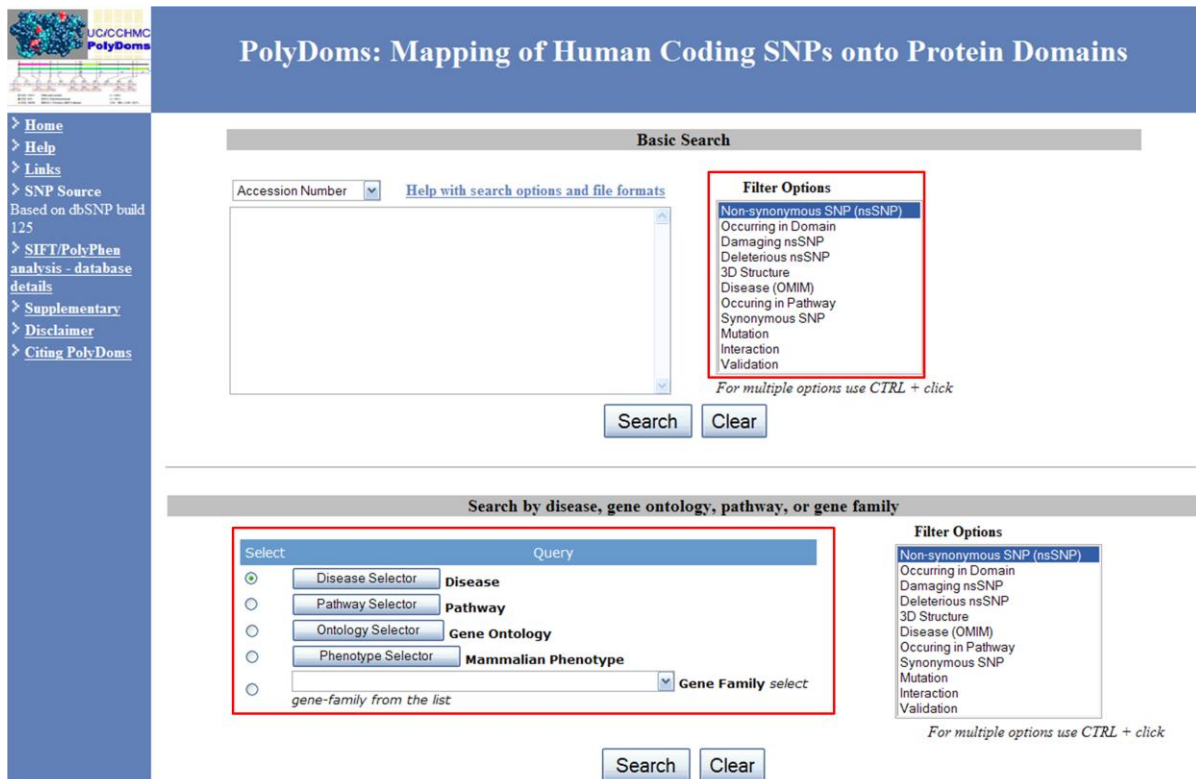
Chapter 5: Annotation of Coding Single Nucleotide Polymorphisms: Using PolyDoms

Description: There is a wide gap between the growing number of reported nsSNPs in human population and the functional consequences of these nsSNPs, and the major challenge lies in distinguishing the functionally significant and potentially disease-related ones from the rest. PolyDoms is based on the hypothesis that if an nsSNP alters the ability of gene products to function normally within the biological pathways or processes, the consequences might either alter disease susceptibility or resistance or result in disease itself or affect the therapeutic regimen. We mapped the coding SNPs (synonymous and non-synonymous) of all the proteins onto their known protein 3D structure and functional domains. We used the coding SNP data from the dbSNP. The database supplemented with a variety of functional implication prediction algorithms like SIFT, PolyPhen, LS-SNP, etc. The web interface supports a variety of queries (GO terms, disease terms, gene families, etc.). Please refer to the “Help” section (<http://info.chmcc.org/help/polydoms/index.html>) on PolyDoms home page (<http://polydoms.cchmc.org>). For any problems or questions or analysis, send a mail to anil.jegga@cchmc.org

Access: Free web-based

Input: Gene symbols, accession numbers, rsSNP IDs, disease terms, GO Terms, etc.

Output: Graphical and results are downloadable as a spreadsheet.



PolyDoms: Mapping of Human Coding SNPs onto Protein Domains

Basic Search

Accession Number [Help with search options and file formats](#)

Filter Options

- Non-synonymous SNP (nsSNP)
- Occurring in Domain
- Damaging nsSNP
- Deleterious nsSNP
- 3D Structure
- Disease (OMIM)
- Occurring in Pathway
- Synonymous SNP
- Mutation
- Interaction
- Validation

For multiple options use CTRL + click

Search by disease, gene ontology, pathway, or gene family

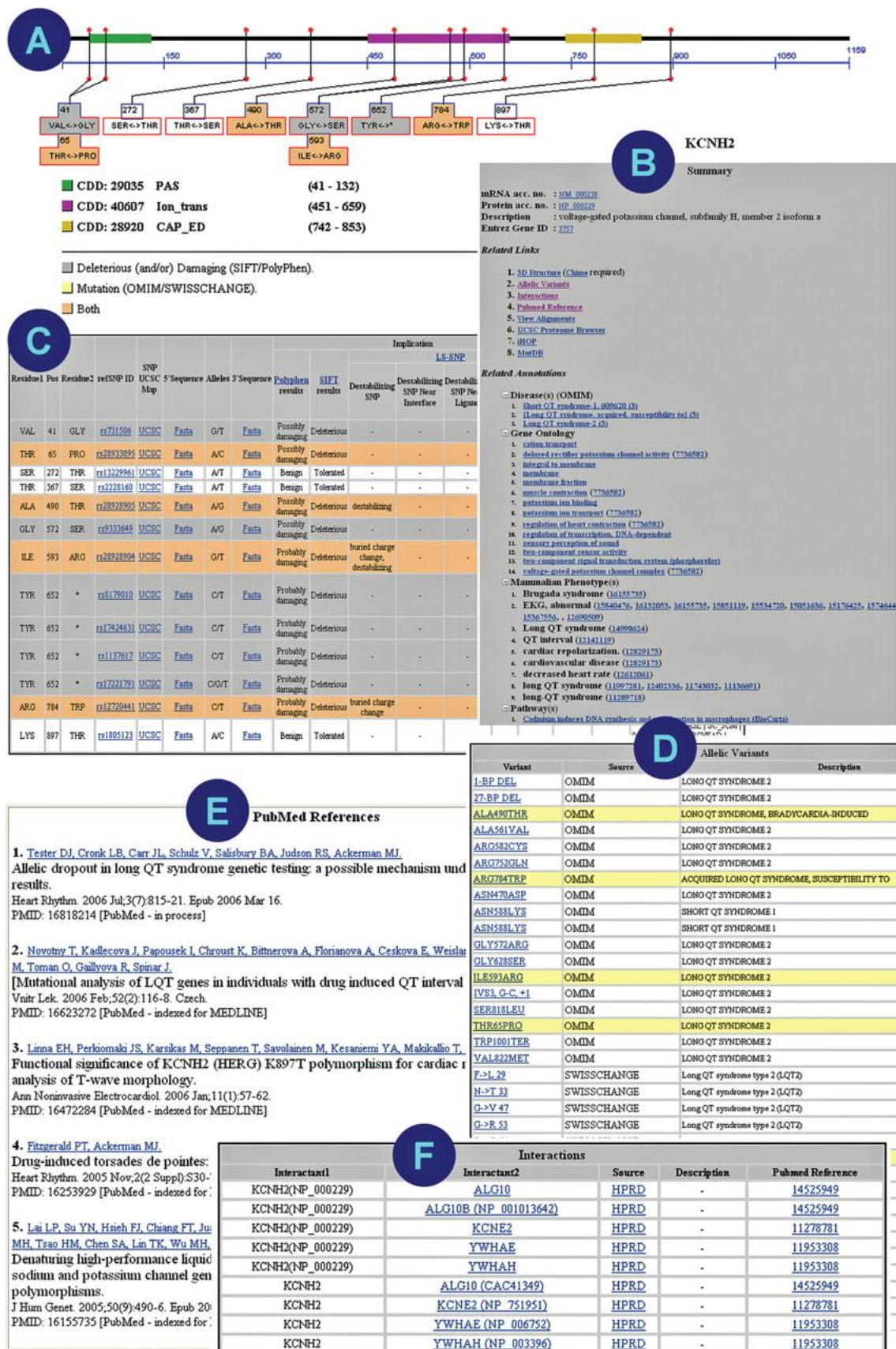
Select ☒ Disease ☐ Pathway ☐ Gene Ontology ☐ Mammalian Phenotype ☐ Gene Family select

gene-family from the list

Filter Options

- Non-synonymous SNP (nsSNP)
- Occurring in Domain
- Damaging nsSNP
- Deleterious nsSNP
- 3D Structure
- Disease (OMIM)
- Occurring in Pathway
- Synonymous SNP
- Mutation
- Interaction
- Validation

For multiple options use CTRL + click



EXERCISE: Transcriptome Analysis and Annotation

1. Using the following two gene lists, identify conserved and non-conserved common binding sites within the upstream 500 bp (*Hint*: when downloading the promoter sequences use 500 bp)

Gene List 1:

ADORA1
AGTRAP
CD37
COL3A1
COL4A2
COL6A1
COL6A3
DCN
E2F4
FN1
LOC374395
LOXL1
LRP3
LRP5
LUM
MMP9
NUMA1
PPARBP
PPARD
PPP2R1A
PSG9
PTGDS
PTPRN
ROM1
SNN
SPARC
THBS2

Gene List 2:

APAF1
BAD
BAX
BCL2
BID
BIRC2
BNIP3L
CASP1
CASP2
CHUK
CYCS

DFFA
DFFB
FADD
FAS
MDM2
MYC
NFKB1
NFKBIA
PRF1
RELA
RIPK1
TNF
TNFRSF10B
TP53
TP73
TRAF1

2. UCSC Browser related:
 - a. Find genes that are predominately expressed in the mouse pancreas, determine the expression pattern of the human ortholog of one such gene and obtain the genomic sequence of the human gene.
 - b. Obtain a list of SNPs in a single gene (*PLG*) using the UCSC Table Browser and annotate them using NCBI's dbSNP batch processor.
 - c. Using one of the gene lists from (1) download all noncoding SNPs occurring within upstream 1 kb region
 - d. Using the same gene list download all coding SNPs. How many of these SNPs are predicted to be deleterious (***Hint***: use PolyDoms once you obtain the list of SNPs. ***Alternate approach***: You can directly use the gene symbols and download the annotated cSNPs including putative deleterious ones from PolyDoms)
3. Using the gene lists (based on published microarray data; Appendix 2), and the applications CisMols, GenomeTrafac and ConciseScanner:
 - a. Find the potential common transcription factor binding site clusters that could be responsible for the co-expression.
 - b. Find additional genome-wide targets for the signature cis-regulatory modules identified by CisMols (***Hint***: use the feature ConciseScanner to find genome-wide additional targets for the shared cis-cluster obtained through CisMols analyzer).
4. Use PolyDoms applications for the following:
 - a. Using one of the gene lists from Appendix 2, annotate all the coding SNPs and find out what SNPs you would consider for designing a diagnostic chip or recommending for re-sequencing?
 - b. What coding SNPs are potentially deleterious for the apoptosis pathway?
 - c. How many proteins are involved in the DNA repair and how many of these have at least one coding nsSNP that occurs in a functional domain and is predicted as deleterious/damaging?

5. Using gene list 2 from (1) above, find out with which other differentially expressed gene lists do they overlap (use MSigDB Annotation or L2L or ToppGene). Are there any gene expression profiling studies where in these genes are down- or up-regulated?
6. Using gene list 1 from (1) above, obtain the mouse orthologs. For the mouse orthologs, download the 3'UTR sequences.

APPENDIX 1: Other Useful bioinformatics resources and tools

1. Bioinformatics Resources: **<http://anil.cchmc.org>**
2. Sequence Manipulation Suite: **<http://anil.chmcc.org/sms/>**
3. RepeatMasker: To mask the repeat elements in a genomic sequence.
<http://www.repeatmasker.org/>
4. PipMaker: To compute alignments of similar regions in DNA sequences.
<http://www.bx.psu.edu/>
5. Exon Mapper: **<http://pbil.univ-lyon1.fr/sim4.php>**
6. EPD Database: Eukaryotic Promoter Database - **<http://www.epd.isb-sib.ch/>**
7. Exon Mapper: **<http://pbil.univ-lyon1.fr/sim4.php>**

APPENDIX 2: Co-expressed gene lists

1: Nitric Oxide. 2002 Nov;7(3):165-86.

A DNA microarray study of nitric oxide-induced genes in mouse hepatocytes: implications for hepatic heme oxygenase-1 expression in ischemia/reperfusion.

Zamora R, Vodovotz Y, Aulak KS, Kim PK, Kane JM 3rd, Alarcon L, Stuehr DJ, Billiar TR.

#NAME inos_10_dn

#DESCRIPTION Ten most-downregulated genes following iNOS induction in hepatocytes

#GENES: CD151, EIF5A, EEF2, CD81, PKM2, ACT6

#NAME inos_10_up

#DESCRIPTION Ten most-upregulated genes following iNOS induction in hepatocytes

#GENES: EED, CSRP1, PCNA, HMOX1, MCM2, CDK2, MCM6, GNB1, TUBB1

2: J Nutr. 2004 Apr;134(4):762-70.

Gene expression profiling in human preadipocytes and adipocytes by microarray analysis.

Urs S, Smith C, Campbell B, Saxton AM, Taylor J, Zhang B, Snoddy J, Jones Voy B, Moustaid-Moussa N.

#NAME adip_human_dn

#DESCRIPTION Down-regulated in primary human adipocytes, versus preadipocytes

#GENES: PPARD, CEBPA, MMP2, SNN, SPARC, COL5A1, DCN, COL3A1, LRP3, COL6A3, PSG9, ATRAP, CD37, ROM1, COL4A2, LUM, PPP2R1A, LRP5, LOX, PTPRN, OKL38, IL18BP, THBS4, FN1, THBS1, LOXL1, COL6A1, ADORA1, MMP9, PPABP, E2F4, PTGDS, THBS2

#NAME adip_human_up

#DESCRIPTION Up-regulated in primary human adipocytes, versus preadipocytes

#GENES: PFKFB3, ABCE1, PLCD1, AGTRL1, CROC4, HSD11B2, AGT, FABP4, DGKG, PTPN21, PTPRZ1, SCD, FABP5, RXRA, SMARCB1, COL1A2, CRYAB, DGAT1, ZNF336, LRP8, CTSG, 3-PAP, APM1, DPT, CAP2, IL22R, SCAP1, USP8, LYPLA1, HPCA, STAT5B, CYB5, E2F5, ALDH6A1, MMP7, LBP, GPD1, GLUL, GPX3, INSR, FXVD1, FAEL2, ALDH1A2, MGST1, MAP4K3, MASP1, ECM2, PTPRS, CEBPD, KCNH2, ATP2B2, ACOX3, SPTBN4, TNFAIP2, LIPE, VN, FABP7, UCP4, LPL, ADFP, PPAR-, E2F1, IGFBP2, CHST1, GDF8, ADORA2B, ATP8A2, ATIP1, LIPC, REQ, PLEK, APOB, TAP1, AMT, PLIN, TFCP2, RXRB

3: Science. 2000 Mar 31;287(5462):2486-92.

Mitotic misregulation and human aging.

Ly DH, Lockhart DJ, Lerner RA, Schultz PG.

#NAME middleage_dn

#DESCRIPTION Downregulated in fibroblasts from middle-age individuals, compared to young

#GENES: CCNB, PLK, FOXM1, KIF11, PTGS2, KIF2C, CENPA, CDC20, H2AFX, KIF23, HMGN2, UBE2C, CCNF, CCNA, CENPF, MYB

#NAME middleage_up

#DESCRIPTION Upregulated in fibroblasts from middle-age individuals, compared to young

#GENES: COL15A1, TNFRSF11B, SERPINB2, COL6A2, IL8, FMOD, MMP12, DPT, CST6, COMP, THBS2, PTGS1, CRYBB2, MMP10, PRSS11

4: Oncogene. 2001 Jun 21;20(28):3674-82.

Distinctive gene expression profiles associated with Hepatitis B virus x protein.

Wu CG, Salvay DM, Forgues M, Valerie K, Farnsworth J, Markin RS, Wang XW.

#NAME hbx_dn

#DESCRIPTION Downregulated by expression of Hepatitis HBx protein in hepatocytes

#GENES: CD4, GSTA4, GLG1, WT1, TGFB1, MAP3K1, IL6, APR-3, TP53, CDKN1A, GSTM5, APC, GAS6

#NAME hbx_up

#DESCRIPTION Upregulated by expression of Hepatitis HBx protein in hepatocytes

#GENES: CCNI, DAD1, GSTM4, AP4B1, CDK4, TYMS, TNFRSF6, MYC, CCND3, PDCD2, PTK9, AP4S1, IGF1R, BCL2L1, TUBG1, TUBA4, TUBG2, VCL, IFNGR1, SLC5A1, MFNG, IFNAR2, CASP4, AP4E1, CDKN3

5: Nat Rev Cancer. 2002 Jan;2(1):38-47.

Hypoxia--a key regulatory factor in tumour growth.

Harris AL.

#NAME hypoxia_review

#DESCRIPTION Genes known to be induced by hypoxia

#GENES: EDN1, PFKP, MMP13, HSF, AK3, BIK, TGM2, P4HA, TEK, CDKN1B, SLC2A3, TF, CCNG2, CD99, SAT, FTL, PFKL, BNIP3, TH, RP1, STC1, HIF2A, PDGFB, VIM, IL8, LDHA, SPP1, CA9, PTGS2, PRPS1, BHLHB2, HK1, IGFBP2, TGFB1, APEX1, TFR3, ALDOA, CCL2, CA12, IL6, SLC2A1, ANGPT2, ACAT, L1CAM, TAGLN, HMOX1, FLT1, ANXA, TGFB3, PGF, IGF2, VEGF, IGFBP1, DDIT3, FOS, LRP8, ENPEP, HK2, G22P1, NOS, ADRA, HIF1A, TGFA, ENO1, PKM2, FGF3, HDAC, CDKN1A, ITGA, BNIP3L, ADM, XRCC5, EDN2, MIF, NFKB1, SERPINE, TXN, IGFBP3, COL5A1, F3, JUN, GAPD, PLAUR, TFF3, EPO, CP, HGF, PGK1

6: Science. 1999 Aug 27;285(5432):1390-3.

Gene expression profile of aging and its retardation by caloric restriction.

Lee CK, Klopp RG, Weindruch R, Prolla TA.

#NAME aged_mouse_muscle_dn

#DESCRIPTION Downregulated in the gastrocnemius muscle of aged adult mice (30-month) vs. young adult (5-month)

#GENES: IL6ST, CALM3, SIN3A, GFER, USP4, ABCB4, COL1A2, PRKCSH, PTPRR, POLA2, PLA2G7, HNRPD, COL1A1, PPP1R2, PRSS15, CLTB, FDFT1, PMP22, PSMB8, MYH2, TST, BMP8B, ADAM28, SRPR, PSMC3, CDC2L2, PPP3CC, S100A10, RAI2, NR2F1, PHOX2A, WNT4

#NAME aged_mouse_muscle_up

#DESCRIPTION Upregulated in the gastrocnemius muscle of aged adult mice (30-month) vs. young adult (5-month)

#GENES: GDF9, ARF5, MFAP5, HSPA6, HSPB1, ETV4, TFAP2B, ISLR, GADD45A, CKMT2, USP53, ATF3, ACTR1B, PBEF1, RAB1A, DCTN1, STARD7, DDX5, TM4SF3, U2AF2, SOX17, RAB21, AP3S2, CDC42, PLAGL1, AMY2B, PRSS11, ZFP90, POU3F2, HINT1, TGFB1I1, TGIF, ARHGDIB

APPENDIX 3: Querying NCBI - Example

Querying the NCBI's Entrez Gene:

For each of these, you can use the “Limits” also.

Purpose	Query	Explanation
find genes mapped to <i>Mus musculus</i> chromosome 16 that have orthologs reported in HomoloGene	Mus musculus[orgn] AND 16[chr] AND gene_homologene[filter]	<ul style="list-style-type: none"> [orgn] is used to restrict to mouse (<i>Mus musculus</i>) to the organism field. Alternatively, you can use the “Limits” form to select mouse only. [chr] is used to restrict '16' to the chromosome field gene_homologene[filter] is used to restrict records to those processed by HomoloGene.
Find genes mapped to human chromosome 16 that have orthologs reported in HomoloGene and also have an OMIM record associated	16[chr] AND gene_homologene[filter] AND gene_OMIM[filter] AND "Homo sapiens"[orgn]	<ul style="list-style-type: none"> Same as previous but an additional filter (OMIM) is used.
find all genes in the NCBI database that are derived from genomes other than mammals and are classified by Gene Ontology to have some association to DNA repair	“dna repair”[go] NOT mammalia[orgn]	<ul style="list-style-type: none"> [go] is used to restrict to the field 'Genome Ontology' Quotes (“dna repair”) are necessary to treat the two words (dna and repair) together [orgn] is used to restrict (as the boolean NOT) to species not classified as

		mammals.
--	--	----------

Querying the NCBI's OMIM:

1. What human genes are related to diabetes? Which of those genes are on chromosome 1?
 - i. enter: diabetes in the search box
 - ii. select Limits
 - iii. check the box for chromosome 1
 - iv. press Go

OR enter the following query in the search box and it will return the same results:

diabetes[All Fields] AND 1[chr]

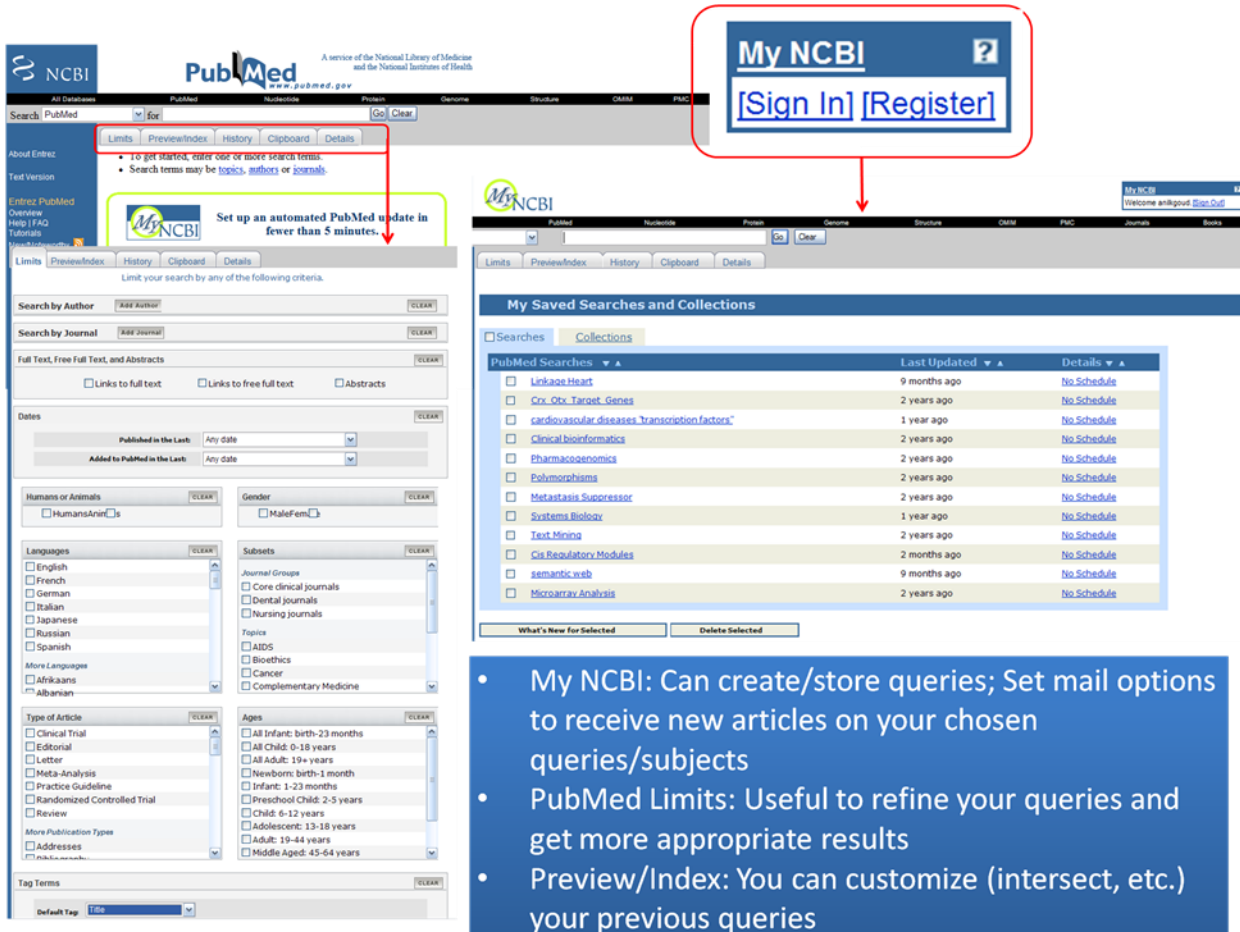
Note that some of the records retrieved will mention hypertension as part of the text of an entry, rather than in the title. That is because Entrez searches All Fields of a record by default. If you would like to limit retrieval only to records that contain the term diabetes in the Title Word field, check the "Search in Field" box for "Title" on the Limits page, in addition to checking the box for Chromosome 1. The query then would be
diabetes[Title] AND 1[chr]

2. List the OMIM entries that describe genes on chromosome 21 and additionally each of which have a clinical synopsis also.

You can do this search in any of these two ways:

- i. Use the Limits page:
 - a. From the OMIM home page, select the Limits option under the search box.
 - b. check the box for chromosome 21.
 - c. Check the box for "clinical synopsis" under the section "Only Records With".
 - d. Press Go.
- ii. Enter the search as a command:
 - a. On the OMIM home page, enter the following in the search box:
 - b. 21[chr] AND "Clinical Synopsis"[prop]
 - c. Press Go.

APPENDIX 4: Coping/Keeping up with literature (PubMed) searches



The screenshot displays the PubMed website interface. At the top, the 'My NCBI' section is highlighted with a red box, containing links for '[Sign In]' and '[Register]'. Below this, the 'Limits' tab is selected, showing various search filters. A red arrow points from the 'Limits' tab to a 'My NCBI' box that says 'Set up an automated PubMed update in fewer than 5 minutes.' Another red arrow points from the 'My NCBI' box to the 'My Saved Searches and Collections' section. This section contains a table of saved searches with columns for 'PubMed Searches', 'Last Updated', and 'Details'.

PubMed Searches	Last Updated	Details
<input type="checkbox"/> Linkage Heart	9 months ago	No Schedule
<input type="checkbox"/> Cox, Otx, Target, Genes	2 years ago	No Schedule
<input type="checkbox"/> cardiovascular diseases, transcription factors	1 year ago	No Schedule
<input type="checkbox"/> Clinical bioinformatics	2 years ago	No Schedule
<input type="checkbox"/> Pharmacogenomics	2 years ago	No Schedule
<input type="checkbox"/> Polymorphisms	2 years ago	No Schedule
<input type="checkbox"/> Metastasis Suppressor	2 years ago	No Schedule
<input type="checkbox"/> Systems Biology	1 year ago	No Schedule
<input type="checkbox"/> Text Mining	2 years ago	No Schedule
<input type="checkbox"/> Cis Regulatory Modules	2 months ago	No Schedule
<input type="checkbox"/> semantic web	9 months ago	No Schedule
<input type="checkbox"/> Microarray Analysis	2 years ago	No Schedule

- My NCBI: Can create/store queries; Set mail options to receive new articles on your chosen queries/subjects
- PubMed Limits: Useful to refine your queries and get more appropriate results
- Preview/Index: You can customize (intersect, etc.) your previous queries